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(54) **CRIMEAN-CONGO HEMORRHAGIC FEVER VIRUS VACCINE**

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CPC **A61K 39/12** (2013.01); **A61K 2039/5254**
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(58) **Field of Classification Search**

None

See application file for complete search history.

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ABSTRACT

The genetically modified hemorrhagic fever virus of this invention possesses a viral ovarian tumor protease with decreased ability to remove ubiquitin (Ub) and ISG15 tags that the human organism uses to label proteins for removal. Unlike complete knockout strains, the modified virus retains enough activity for replication in a human cell line. This creates an immunogenic and non-pathogenic virus that can be used as an effective live vaccine agent.

17 Claims, 9 Drawing Sheets

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Figure 1

Crimean-Congo hemorrhagic fever (CCHF)
(-) ssRNA Viral Genome

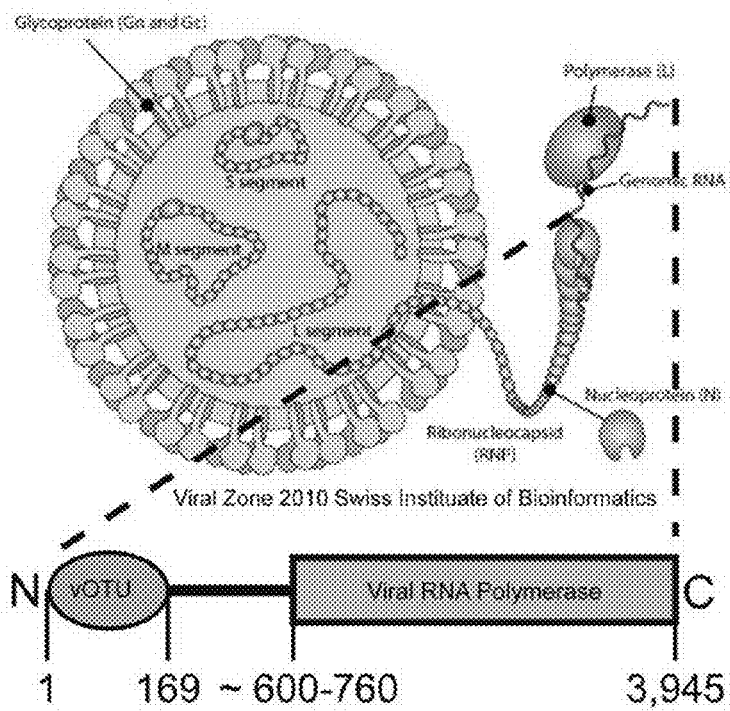


Figure 2

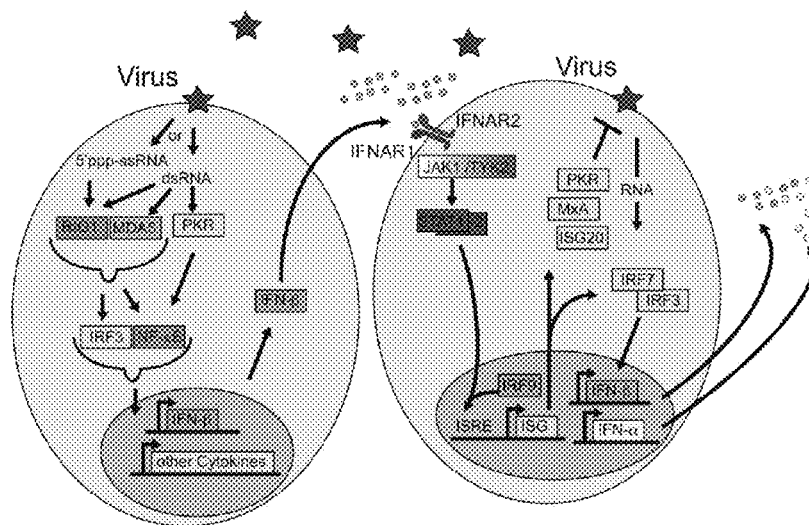


Figure 3

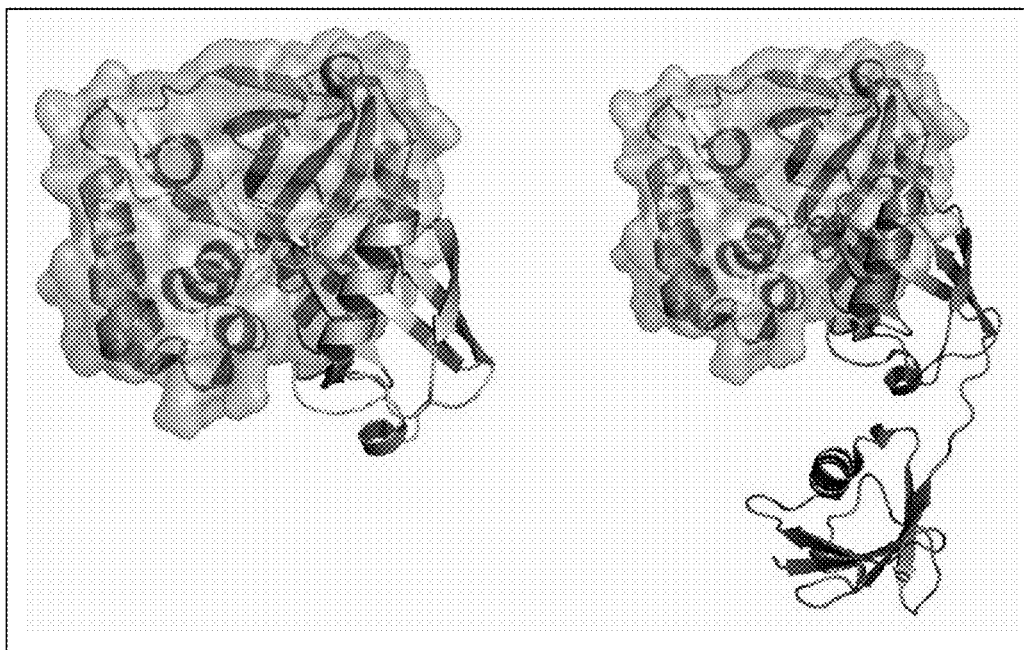


Figure 4

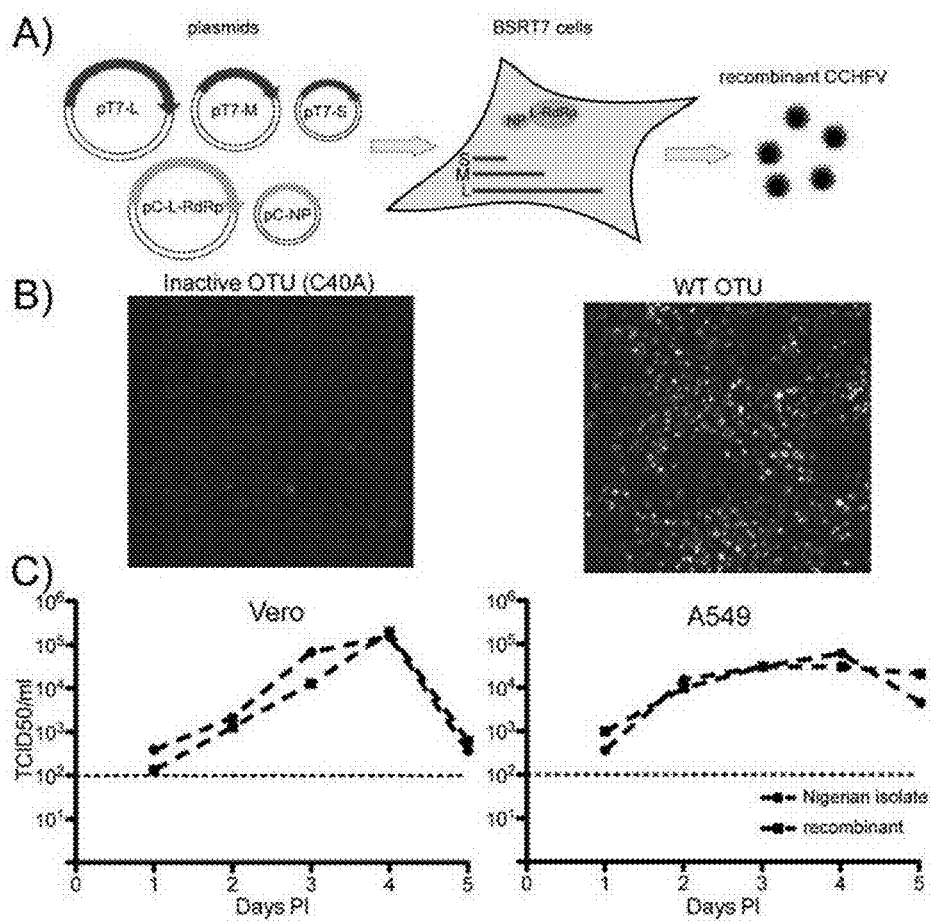


Figure 5

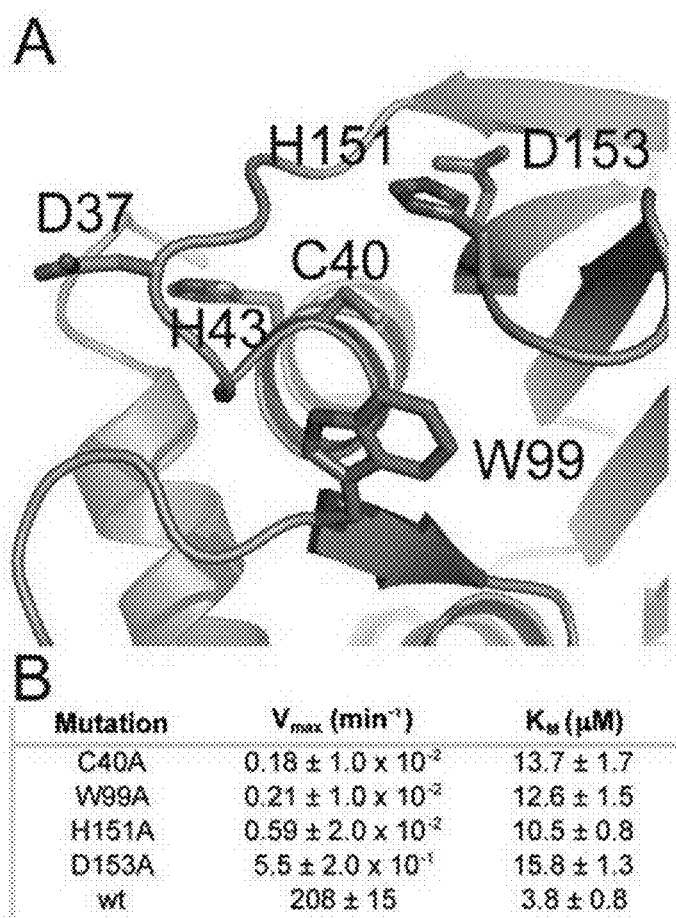


Figure 6

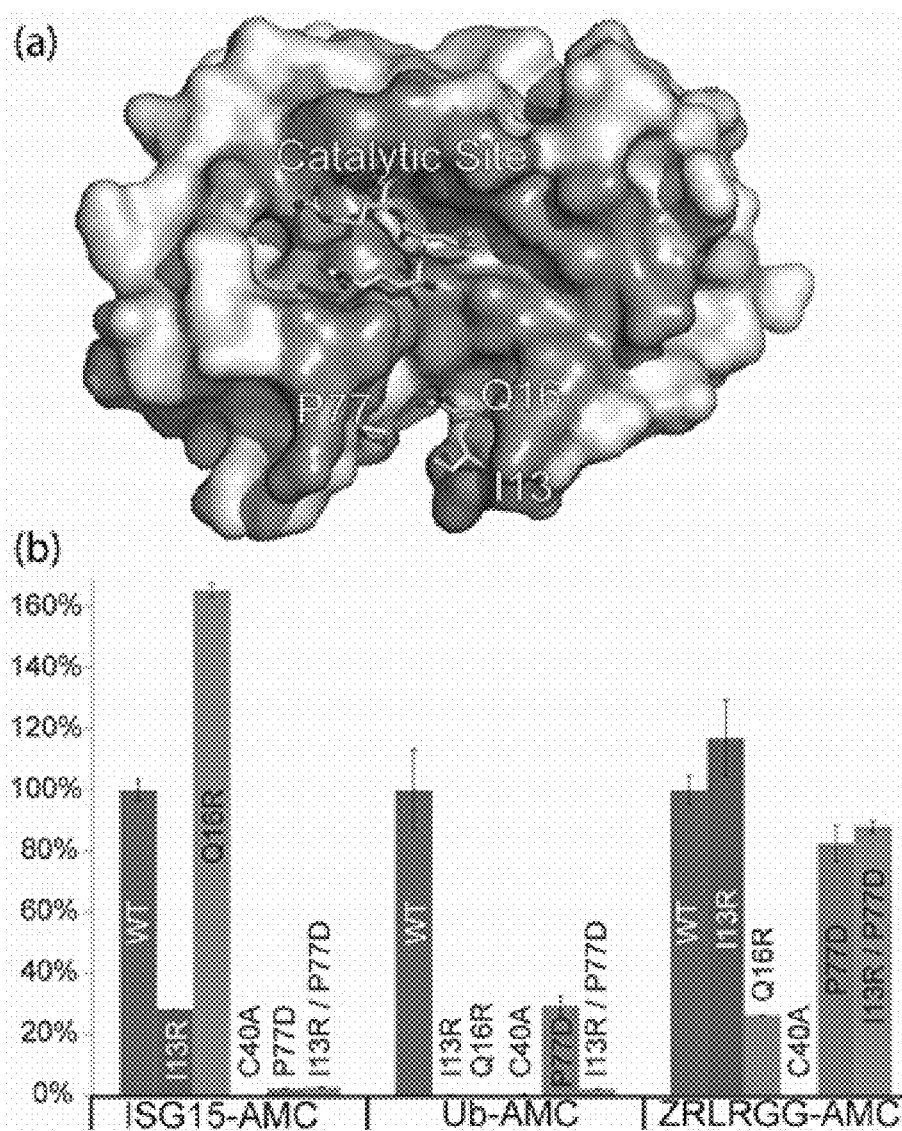


Figure 7

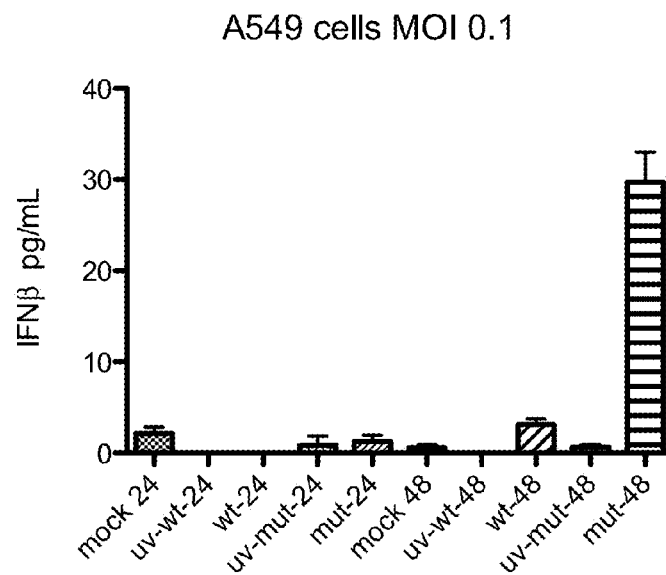


Figure 8

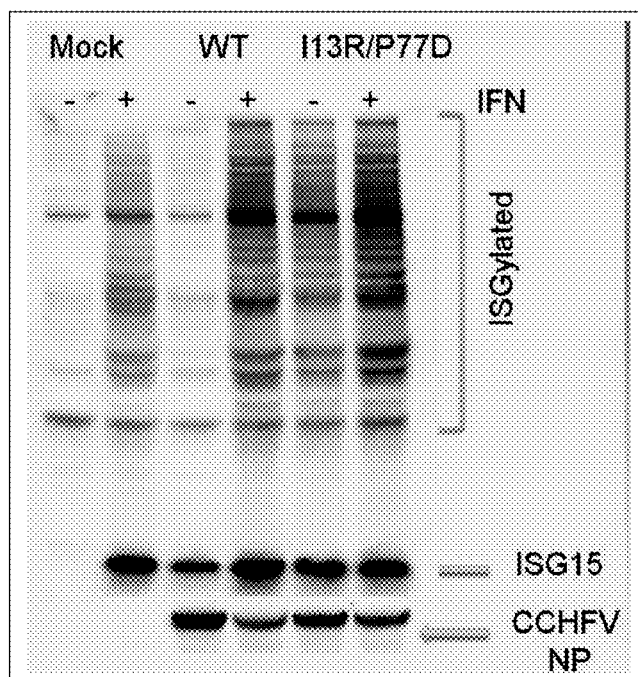


Figure 9

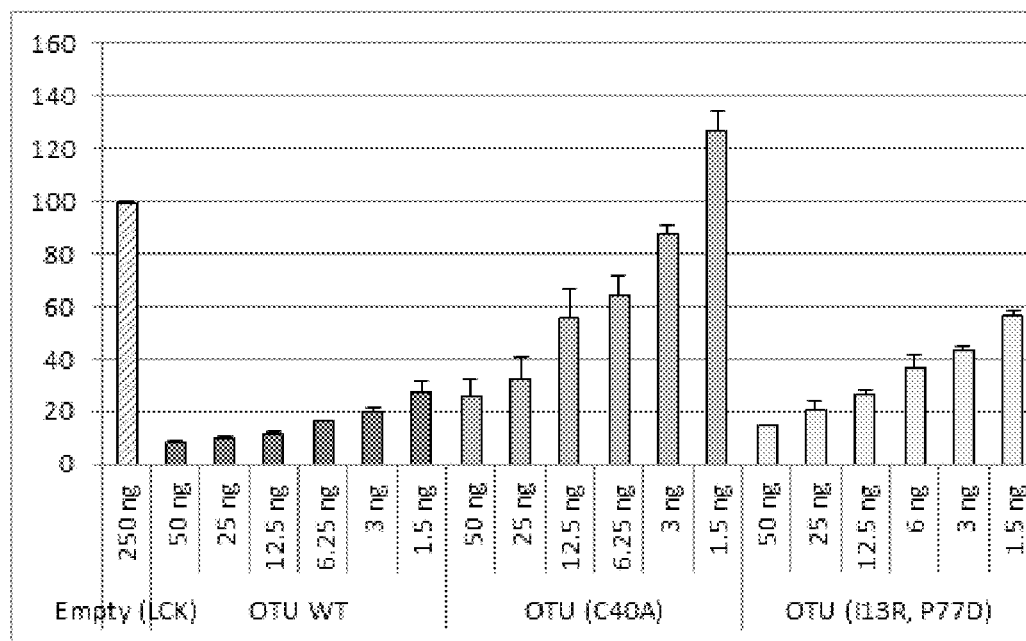


Figure 10

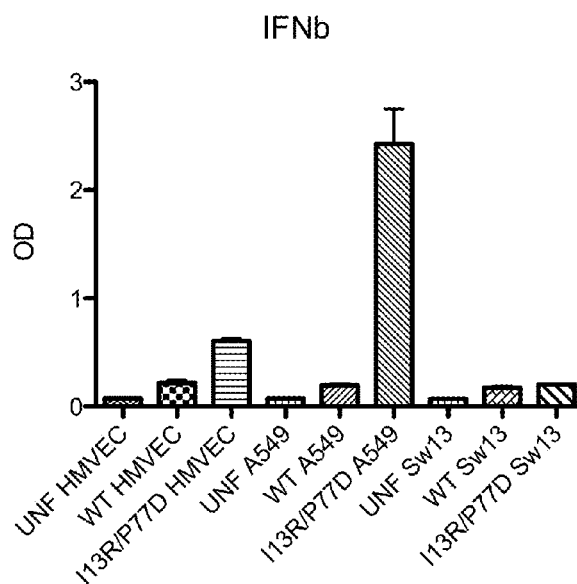


Figure 11

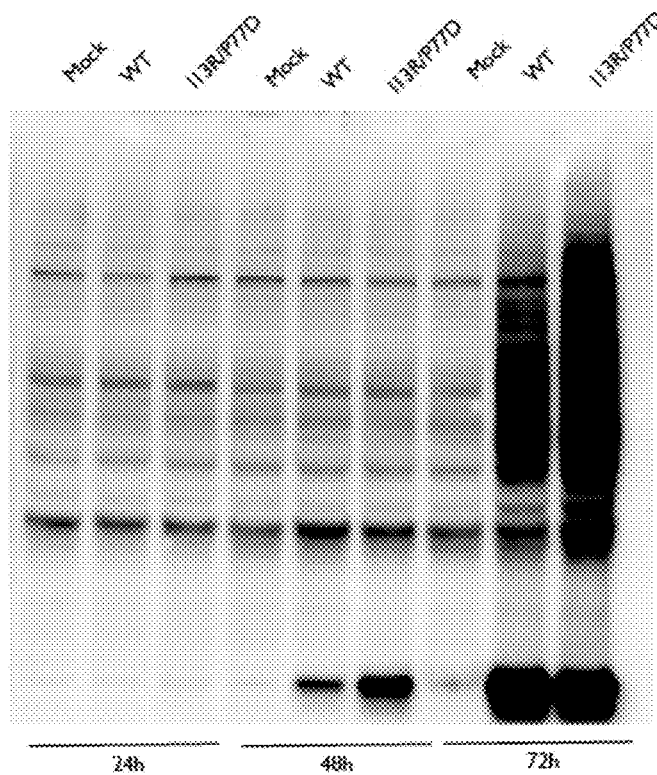


Figure 12

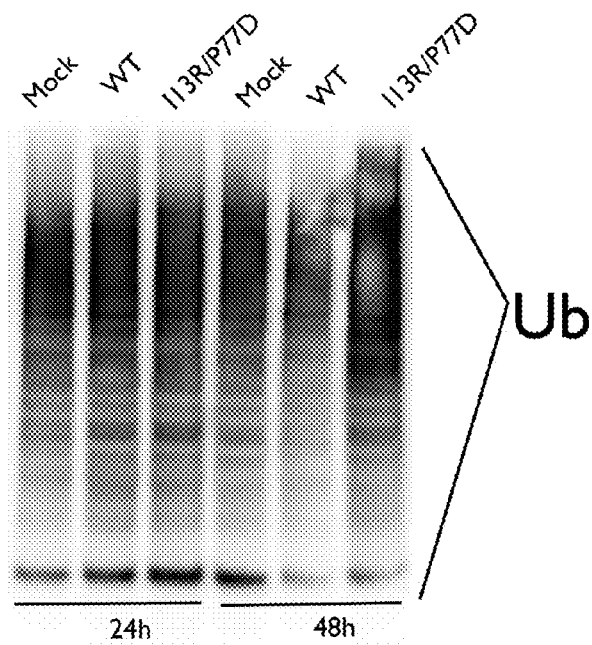


Figure 13

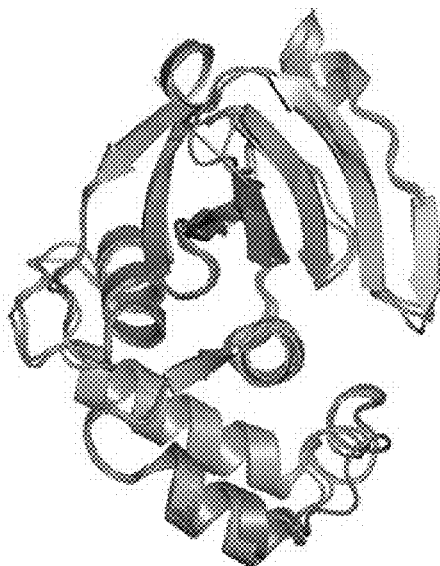


Figure 14

10	*	*	80		
WTQVIDG		EEPEAKL		Afg09-2990	CCHV strains
WTQVIDS		EEPEAKL		Matin	
WTQVIAG		EEPEAKL		Oman	
WTQVMAG		EEPEAKL		Baghdad	
WTQVIAG		EEPEAKL		UG3010	
WTQVIAS		EEPEAKL		Turkey	
WTQVIAG		EEPEARL		IbAr10200	
WTQVIAG		EEPEAKL		SPU128-81	
WTQVLAG		DEPEAKL		AP92Greece	
10	*	*	80		
WERVVDE		TEPEAVGT		DUGV	other nairoviruses
WTQVIAG		EEPEARLV		CCHFV	
WEEVPG		EEPEAKGL		NSDV	
WENIEG		TEPEAIGL		ERVEV	
WDSVSDI		TEPEAAAT		HAZV	

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CRIMEAN-CONGO HEMORRHAGIC FEVER VIRUS VACCINE

REFERENCE TO RELATED APPLICATIONS

This application claims the priority benefit of U.S. provisional application 61/683,132, filed Aug. 14, 2012. The priority application is hereby incorporated herein by reference in its entirety for all purposes.

GOVERNMENT SUPPORT

This invention was made in part with government support under NIH 1R03AI092249-01 awarded by the National Institutes of Health. The government has certain rights in the invention.

FIELD OF THE INVENTION

This application relates generally to the field of viral disease, prophylaxis, and vaccination. More specifically, it provides a virus vaccine modeled on the etiologic agent for Crimean-Congo hemorrhagic fever. It was produced by reducing the deubiquinating and deISGylating activities from the viral OTU protease.

BACKGROUND

Crimean-Congo hemorrhagic fever (CCHF) is a widespread tick-borne viral disease that can affect humans. It is a member of the Bunyaviridae family of RNA viruses. Clinical disease is rare in infected mammals, but it is commonly severe in infected humans. Outbreaks of illness are usually attributable to handling infected animals or people.

The causative organism is found in Asia, Eastern Europe, the Middle East, a belt across central Africa and South Africa and Madagascar. The main environmental reservoir and vector for the virus is hard ticks. Ticks carry the virus to domestic animal stock. Sheep, goats and cattle can develop viremia, but tend not to fall ill. Tick species that have been identified as infected with this virus include *Argas reflexus*, *Hyalomma anatolicum*, *Hyalomma detritum*, *Hyalomma marginatum* and *Rhipicephalus sanguineus*.

The onset of CCHF is sudden, with initial signs and symptoms including headache, high fever, back pain, joint pain, stomach pain, and vomiting. Red eyes, a flushed face, a red throat, and petechiae (red spots) on the palate are common. Symptoms may also include jaundice, and in severe cases, changes in mood and sensory perception. As the illness progresses, large areas of severe bruising, severe nosebleeds, and uncontrolled bleeding at injection sites can be seen, beginning on about the fourth day of illness and lasting for about two weeks.

Animal herders, livestock workers, and slaughterhouses in endemic areas are at risk of CCHF. Healthcare workers in endemic areas are at risk of infection through unprotected contact with infectious blood and body fluids. Individuals and international travelers with contact to livestock in endemic regions may also be exposed. In documented outbreaks of CCHF, fatality rates in hospitalized patients have ranged from 5% to as high as 80%.

Previous attempts to develop preventative treatment are as follows. In a USSR/Bulgarian CCHF vaccine developed in 1974 comprised an inactivated antigen from CCHF virus strain V42/81. It was generated from suckling mouse brain preparations, and so is unsuitable for FDA approval in the

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U.S. There is also a recombinantly produced construct comprising G1 (Gc), or G2 (Gn) glycoprotein ectodomains or portions thereof. However, no study exists to suggest any efficacy for this approach. Full effectiveness of this construct may be limited to the specific strain where the selected glycoproteins originated. There is no established virus-specific treatment. Ribavirin is thought to be effective in vitro, and has been used in human subjects during outbreaks. There are conflicting reports as to effectiveness, with the more recent ones showing limited to no effectiveness against CCHF virus in vivo.

The Department of Defense views CCHF virus as a potential threat to the U.S. armed forces when operating in countries endemic to the virus. These geographical locations include but are not limited to Afghanistan, Pakistan, and the Middle East. The need for preventative treatment of was underscored by death of a U.S. soldier from CCHF viral infection in 2009.

SUMMARY OF THE INVENTION

This invention provides a genetically modified hemorrhagic fever virus that has a viral ovarian tumor protease with decreased ability to remove ubiquitin (Ub) and ISG15 tags from proteins in the cells it infects. Unlike complete knockout strains, the modified virus retains enough activity for replication in a human cell line. This creates an immunogenic and non-pathogenic virus that can be used as an effective live vaccine agent.

One aspect of this invention is a pharmaceutical composition effective in eliciting a specific immune response, that is capable of replication in human cells, but that has been recombinantly altered to have decreased deubiquinating activity or decreased deISGylating activity while maintaining protease activity. Any hemorrhagic fever virus, nairovirus, or a member of the Bunyaviridae family of RNA viruses can be tested for suitability of this invention. A non-limiting example is Crimean-Congo hemorrhagic fever (CCHF) virus, which is used to illustrate the more general aspects of the invention in this disclosure.

Immunogenic compositions of this type can be recombinantly altered to have decreased deubiquitinating activity and/or decreased deISGylating activity. Typically, a lower level of deubiquitinating activity and a lower level of deISGylating activity remain in the mutant virus so that the virus can replicate in a suitable host cell: for example, less than 10%, 5%, or 2% of the activity of either or both deubiquitinating activity and deISGylating activity.

By way of illustration, the immunogenic composition may be modified at position 13, position 77, or both position 13 and 77 of the L-protein. Position 13 of the L-protein may be changed to arginine; position 77 may be changed to aspartic acid. The immunogenic composition may further comprise an adjuvant. After modification, the vOTU protein may have no ability or a reduced ability to inhibit expression of interferon β .

A related aspect of the invention is a recombinant CCHF virus that has been modified to have both decreased deubiquitinating activity and decreased deISGylating activity, and that is capable of replication in human cells. The invention includes other viruses that have been recombinantly engineered or mutated to reduce deubiquitinating and deISGylating activity. This includes Dugbe virus (DUGV), Hazara (HAZV), Nairobi sheep disease virus (NSDV), Ganjam virus (GANV), or any virus that causes febrile illness of varying severity in humans, pets, and agricultural animals.

Included in the invention are host cells transfected with an engineered virus having one or more of the properties indicated above.

Another aspect of the invention are methods for eliciting a specific immune response and/or for preventing or treating hemorrhagic fever, using a recombinant virus or immunogenic composition. Also provided are methods for preparing a commercial product wherein a composition or virus is packaged with information on use.

This invention also provides a method of developing an immunogenic but substantially non-pathogenic hemorrhagic fever virus. A host cell is transfected with the genome of a wild-type hemorrhagic fever virus along with genetic material comprised of a codon optimized L-protein. The genome has one or more genetic alterations introduced before transfection. Viral particles are recovered, and then tested and selected for decreased deubiquitinating activity and/or decreased deISGylating activity. The method may entail transfecting the host cell with the L, M, and S gene sectors in separate vectors.

Another aspect of the invention is a method for preparing a commercial product. A vaccine or pharmaceutical composition of the invention is packaged with information on how to use the product for eliciting an immune response or for preventing or treating hemorrhagic fever.

Other aspects of the invention will be apparent from the description that follows.

DESCRIPTION OF THE DRAWINGS

FIG. 1 depicts structural features of CCHF virus and otherairovirus related diseases.

FIG. 2 illustrates the molecular pathway and modulation of the innate interferon (IFN) type 1 mediated immune response.

FIG. 3 is a three-dimensional representation of the ubiquitin and ISG15 proteins docking with the Viral Ovarian Tumor Domain Protease (vOTU) of CCHF virus.

FIG. 4 shows the reverse genetics method developed to produce recombinant CCHF virus in T7 RNA pol. expressing cells.

FIG. 5 depicts the active site of vOTU as a three-dimensional rendering.

FIG. 6(a) shows the residues selected for mutation as part of the three-dimensional structure of vOTU. FIG. 6(b) presents data showing disruption of the vOTU deubiquitinating and deISGylating activities in vitro

FIG. 7 shows results of an assay for interferon (IFN) beta (β) in cells infected with CCHF virus wild type (WT) and the selected mutant.

FIG. 8 shows results of monitoring ISGylation of wild type (wt) CCHF virus and reverse genetically produced CCHF virus containing the I13R/P77D mutation.

FIG. 9 shows data comparing the ability of the engineered virus with a totally inactive mutant virus (C40A) to inhibit production of interferon beta.

FIG. 10 shows a test for interferon β production in lung carcinoma A549 cells

FIG. 11 shows a Western blot testing human ISG15 activity.

FIG. 12 shows a Western blot of the total levels of cellular protein ubiquitination in cells following infection.

FIG. 13 compares the three-dimensional structure of the vOTU protein in CCHF and Dugbeairoviruses.

FIG. 14 shows that residues P77 and I13 are highly conserved amongst strains of CCHFV (SEQ ID NOS:4-12)

(top) and otherairoviruses (SEQ ID NOS:13, 14, 7 and 15-21) (bottom), particularly those known to cause human disease.

DETAILED DESCRIPTION

Context

The Crimean-Congo hemorrhagic fever (CCHF) virus is a member of the genus *Nairovirus*, family Bunyaviridae. The negative sense RNA genome is composed of three segments—Small (S), Middle (M) and Large (L). The L segment is 11-14.4 kilobases in length while the M and S segments are 4.4-6.3 and 1.7-2.1 kilobases long respectively. The L segment encodes the RNA polymerase; the M segment encodes the envelope proteins (Gc and Gn); and the S segment encodes the nucleocapsid protein. The envelope protein is initially translated as a glycoprotein precursor which is then cleaved into the mature structural glycoprotein products (Gn and Gc) and non-structural glycoproteins.

CCHFV is not the onlyairovirus that causes human disease. Dugbe virus (DUGV), Hazara (HAZV), Nairobi sheep disease virus (NSDV), and Ganjam virus (GANV) all result in varying severity of febrile illness and are located in a subset of countries within the CCHFV endemic region. Additionally, infection with NSDV and the closely related GANV in sheep negatively impacts local economies through high livestock mortality and limiting of trade with the affected areas. ERVV, found in Germany, France, Netherlands, and the Czech Republic, is increasingly implicated as the causative agent of severe headaches, known as thunderclap headaches, which result from subarachnoid hemorrhages in humans.

Further information about these viruses is provided by Yadav, P. D. et al., *Infect Genet Evol* 11, 1111-1120, 2011; Dilcher, M. et al., *Virus Genes*, Aug. 7, 2012; Schwedt, T. J. et al., *Lancet Neurol* 5, 621-631, 2006; and Woessner, R. et al., *Infection* 28, 164-166, 2000. Further information on the CCHF virus as a model for other viruses in this family, including its structure, and biology, can be found in the following publications: Khan A, et al. *Viral Hemorrhagic Fevers*. Seminars in Pediatric Infectious Diseases. Philadelphia: WB Saunders Co., 1997; 8 (suppl 1):64-73; Peters C J. *Viral Hemorrhagic Fevers*. Viral Pathogenesis. New York: Lippincott-Raven Publishers, 1997:779-794.

Ubiquitin is a small intracellular protein that becomes conjugated to and marks proteins for destruction or for transport to particular compartments inside the cell. Ubiquitination is an enzymatic post-translational modification process in which the carboxylic acid of the terminal glycine in activated ubiquitin is catalyzed to form an amide bond to the epsilon amine of the lysine in the modified protein.

Interferon-induced 17 kDa protein ISG15 is a protein that is expressed in response to interferon. ISG15 shares several properties with other ubiquitin-like molecules. Its activity is tightly regulated by specific signaling pathways that have a role in innate immunity. It also has cytokine activity. The mechanism of ISGylation is similar to that of ubiquitination.

Wild-type hemorrhagic fever viruses have both deubiquitinating and deISGylating activity to reverse labeling by ubiquitin and ISG15 as part of its arsenal of weaponry that it brings to bear upon infection of the host.

Overview of the Invention

It has now been discovered that impairment but not elimination of the ability of the virus to remove ubiquitin (Ub) and ISG15 tags creates an immunogenic and non-pathogenic virus that can be used as an effective live vaccine agent.

Post-translational modification of host proteins by ubiquitin (Ub) and Ub-like interferon stimulated gene product 15 (ISG15) known as ubiquitination and ISGylation, respectively, is a way that the human organism tags proteins for removal and degradation. Ubiquitin is a small regulatory protein found in almost all tissues that directs protein recycling by attaching to proteins and labeling them for destruction. The ubiquitin tag directs proteins to the proteasome, which is a large protein complex in the cell that degrades and recycles unneeded proteins. Interferon-induced 17 kDa protein is a protein that in humans is encoded by the ISG15 gene. ISG15 shares several common properties with other ubiquitin-like molecules (UBLs), but its activity is tightly regulated by specific signaling pathways that have a role in innate immunity. Upon interferon treatment, ISG15 can be detected in both free and conjugated forms, and is secreted from monocytes and lymphocytes where it can function as a cytokine.

CCHF virus and all other nairoviruses including Dugbe virus (DUGV), Hazara (HAZV), Nairobi sheep disease virus (NSDV), and Ganjam virus (GANV) possesses a protease (specifically, the viral ovarian tumor domain protease) that performs deubiquitination and deISGylation functions. This enables the virus to evade the human immune response by down-regulating immunological functions such as expression of interferon as well as other antiviral effector and signaling proteins. However, complete loss of function of this protease results in the inability of CCHF virus and likely other nairoviruses to replicate. This prevents viruses that have been genetically modified to eliminate these activities entirely from being useful as a self-propagating vaccine agent.

The genetically modified virus of this invention possesses a viral ovarian tumor protease with significantly less deubiquitination and deISGylation activity, while still retaining enough activity for virus production in a human cell line. The modified virus will not efficiently evade the human immune response, but will generate a level of immunity in the host that protects against future infection by a wild-type virus.

Development of Modified Strains of Virus

The invention described in this disclosure was developed using recombinantly sourced Crimean-Congo hemorrhagic fever virus as a model. The model CCHF virus strain was recovered from hamster cell line (BSR/T7) and propagated in human cell lines. Selective mutations were generated that result in the simultaneous ablation of the greater than 95% deubiquitinating and deISGylating in vitro activity of virus's viral ovarian tumor domain protease.

Reverse genetic derived infectious Crimean-Congo hemorrhagic fever virus strain IbAr10200 may be achieved by first cloning the originating virus's cDNA, or by completing gene synthesis, of the complete segments (S, M and L). The S, M, and L segments were cloned in the pT7 vector between a T7 promoter, to drive the transcription of Crimean-Congo hemorrhagic fever virus complementary genome RNA copies, and a hepatitis D ribozyme, to obtain authentic 3' termini. The vectors were transfected into BSR/T7 cells to obtain recombinant RNA genome matching the cloned sequence. Complementation of the with mammalian expression vectors pCAGGS encoding a human codon optimized L-protein (pC-L) and wild-type N protein (pC-N) is used to obtain recombinant virus.

Details were as follows: Wild recombinant CCHF virus was rescued by transfecting a 10 cm² well of subconfluent BSR/T7/5 cells with 2.5 µg pT7-S, 1 µg pT7-M, 1 µg pT7-L, 0.66 µg of pC-N and 0.33 µg of human codon optimized

pC-L mixed with 11 µL of Mirus LT1™ transfection reagent (Mirus Bio LLC, Madison, Wis.) in OPTI-MEM™ media. All viruses recovered were harvested from cell supernatants four days post transfection and amplified in SW13 cells.

A CCHF vOTU expression construct was obtained by use of an *Escherichia coli* BL21 codon-optimized synthesis of the first 169 amino acids from the L protein in CCHF virus (GenBank accession no. AAQ98866.2) by Biobasic, Inc. Along with the vOTU portion of the L protein, six histidine codons and a stop codon were added to the gene in order to provide a C terminus histidine tag. The resulting gene was incorporated into a pET11a plasmid using NdeI and BamHI restriction sites. Site directed mutagenesis of the construct was performed using a QuikChange™ kit. Successful mutations were confirmed by sequencing performed by GenScript™. The mutated constructs were then transformed into BL21(DE3) cells, and were grown at 37° C. in 6 L of LB broth containing 100 µg/mL of ampicillin until the optical density at 600 nm reached 0.6. Expression of wild type (WT) or mutant CCHF vOTU was induced by the addition of IPTG to a final concentration of 0.8 mM. The culture was further grown for 4 hrs at 37° C. and then centrifuged at 6,000×g for 10 minutes. Cells were collected and stored at -80° C. until use. vOTUs were purified according to a standard protocol and assayed for activity.

CCHFV L amino acid positions 13 and 77 were mutated to isoleucine and aspartic acid and replaced the wild type pT7-L vector in the transfection plasmid mix. Four days following the transfection, immunoreactive foci can be detected and recovery of infectious recombinant Crimean-Congo hemorrhagic fever virus was confirmed by passing the transfection supernatants to SW13 cells. Three days later, cytopathic effect can be evident and Crimean-Congo hemorrhagic fever virus antigens can be detected throughout a cell monolayer.

Mutation of the 13th and 77th amino acid positions within their L-protein to isoleucine and aspartic acid respectively create a mutant lacking significant Ub and ISG15 activity, while maintaining activity to cleave a peptide. Aberration of complete activity of the viral ovarian tumor domain protease that is located in 1-169 amino acids of the L-protein by a mutation of position 40 from cysteine to alanine results in no recombinant virus.

Mutation of position 77 of the L-protein to aspartic acid results in the viral ovarian tumor protease of Crimean-Congo hemorrhagic fever virus strains is necessary to disrupt a hydrophobic interaction between it and human interferon stimulated gene product 15. This significantly reduces the ability of the viral ovarian tumor protease to recognize stimulated gene product 15.

To remove deubiquitinating activity, mutation of position 13 of the L-protein to arginine interferes through charge repulsion with an arginine at position 42 in ubiquitin and a tryptophan at position 123 in interferon stimulated gene product 15. This double mutation reduces deubiquitinating and deISGylating activities to 2% and 3% that of wild-type viral ovarian tumor protease, respectively, while maintaining catalytic activity greater than 88% that of wild-type viral ovarian tumor protease in vitro.

To construct the recombinant virus, the gene encoding native L-protein is altered at position 77 and position 13 of the amino acid sequence to delete the residue or substitute a residue or plurality of residues that is different from the native sequence. For example, the amino acid substitution at position 13 in the L-protein could be lysine or histidine. The amino acid substitution at position 77 in the L-protein could be other amino acids with a polar or charged side chain.

The I13R/P77D double mutation eliminates CCHF virus's viral ovarian tumor (vOTU) domain protease from performing deubiquitinating and deISGylating activity, but it still allows the virus to replicate. The CCHF virus with the I13R/P77D changes maintains one or more critical innate immunity biomarkers.

Illustrations

FIG. 1 depicts structural features of CCHF virus and the etiologic agent for other nairovirus related diseases. Rift Valley Fever Virus possesses an S-segment encoded NSs virulence factor, which allows for immune system evasion. Removal of NSs results in virus that does not effectively evade immune system. Crimean-Congo hemorrhagic fever (CCHF) virus does not encode a NSs factor, but it does have a vOTU (Viral Ovarian Tumor Domain Protease: see G C Capodagli et al., J Virol. 2011 April; 85(7): 3621-3630).

FIG. 2 illustrates the molecular pathway and modulation of the innate interferon (IFN) type 1 mediated immune response.

FIG. 3 is a three-dimensional representation of the ubiquitin and ISG15 proteins docking with the vOTU protein of CCHF virus, developed from the crystal structure of CCHF virus determined by Capodagli et al. supra.

FIGS. 4 and 5 show the reverse genetics CCHF virus system used for developing the invention. FIG. 4(A) shows the method developed to produce recombinant CCHF virus in T7 RNA pol. expressing cells. The solid arrows depict the genome RNA produce by the T7 ("pT7"), and viral proteins supporting the initial genome replication ("pC"). The panels below show immunofluorescence detection of CCHF virus produced by reverse genetics.

FIG. 5 depicts the active site of vOTU. (A) is a three-dimensional rendering of vOTU's active site, showing secondary structures, helices, and loops. (B) Mono-Ub Km and Vmax constants determined for catalytic triad vOTU mutants.

FIG. 6 is taken from the development of CCHF virus vOTU-I13R/P77D. FIG. 6(a) shows data from disruption of the vOTU deubiquitinating and deISGylating activities in vitro. The CCHF virus vOTU is shown with the residues which comprise the complete vOTU/Ub binding interface. Residues Q16 and I13 were selected to disrupt the binding of Ub through site directed mutagenesis. P77 was selected to disrupt binding of ISG15 through mutagenesis. The peptide RLRGG represents the C-terminal tail of Ub and ISG15. FIG. 6(b) shows data from disruption of the vOTU deubiquitinating and deISGylating activities in vitro.

In FIG. 7 interferon (IFN) β was monitored from immunocompetent A549 cells that were infected with UV inactivated wt CCHF virus (uv-wt), wt CCHF virus (wt), I13R/P77D CCHF virus (mut). Upon infection, bsr7 cells are not interferon producing cells, whereas A549 are. 24 and 48 denotes the time points for surveying IFN β production. For uv-wt, the virus is inactivated and incapable of infection, thus no IFN β production. Wild type CCHF virus has a functioning vOTU that suppresses IFN β production. However, I13R/P77D renders CCHF virus's vOTU unable of performing that function resulting in a significantly observable change in IFN β level over 48 hours.

FIG. 8 shows results of monitoring ISGylation of wild type (wt) CCHF virus and reverse genetically produced CCHF virus containing the I13R/P77D mutation within CCHF virus's vOTU. ISG15 antibodies were used to highlight proteins that have been ISGylated within A549 cells upon mock infection or infection by wt CCHF virus or I13R/P77D CCHF virus. Antiserum specific for CCHF nucleocapsid was used as a control to confirm CCHF virus

infection. As mock infection contains no virus, no significant ISGylation occurs. Infection of wt CCHF virus reduces the ISGylation to mock levels where as the CCHF virus containing the I13R/P77D mutation can't reduce intracellular ISGylation levels. The (+) columns denote addition of exogenous interferon to probe to evaluate the extent of CCHF virus vOTU activity.

FIG. 9 shows the reduced ability of transfected I13R/P77D at suppressing the transcription activation of an interferon β promoter relative to a totally inactive mutant (C40A) and wild type (WT) vOTU in human embryonic kidney 293 cells. FIG. 10 shows that I13R/P77D CCHFV lack of ability to suppress human immunity (as measured by interferon β production) is lung carcinoma A549 cells and primary culture of human microvascular endothelial cells (HMVEC).

FIG. 11 shows a Western blot for human ISG15 in A549 cells infected with wt CCHFV or I13R/P77D. Mock-infected lanes are also included. Cells infected with I13R/P77D have a significantly higher concentration of ISG15 conjugated proteins (the proteases substrate), then WT (wild type). The mock infected cells have no virus in them, and establish a basal level for ISG15 activity in this assay. FIG. 12 shows a Western blot of the total levels of cellular protein ubiquitination in A549 cells following WT and I13R/P77D infection. This indicates that ubiquitination level is enhanced only by the I13R/P77D infection after 48 h.

FIG. 13 shows the crystal structure of CCHF vOTU (virus ovarian tumor domain) overlaid with that the recently elucidated vOTU from the Dugbe nairovirus. This illustrates that nairovirus vOTUs have a conserved 3-D structure placing I13R and P77D in the same location throughout nairoviruses vOTUs. Similarly, FIG. 14 shows that P77D and I13R are highly conserved amongst strains of CCHFV (top) and other nairoviruses (bottom), particularly those known to cause human disease, the I13 and P77 amino acid sites are conserved.

This shows the general applicability of this invention to create recombinant forms of any one of these viruses and other homologs to have decreased deubiquitinating and decreased deISGylating activity while maintaining protease activity.

Testing and Commercial Use for Immunization and Treatment

Once a virus according to this invention has been generated and tested in tissue culture, its ability to elicit an immune response and/or prevent viral infection can be tested in a suitable animal model. Suckling mice is a suitable system to test the benefits of the vaccine. For proof of concept, a homologous nairovirus can be used. For NSDV (Nairobi Sheep Disease Virus), sheep are the ideal and easiest test model, since it is often fatal in sheep. For Erve virus, wild-type mouse models can be used. For Dugbe, Hazara, or Erve virus, suckling mice is an accepted model for the safety and efficacy of the vaccine as their immune system is immature.

In any of these models, a suitable end point would be protection, reduced fever, reduced duration of infection, or at least prolonged survival. Blood samples are taken before the testing and periodically after administration to measure antibody response, cellular response, and virus inhibition. An increase in any one or more of these responses is expected to correlate with clinical efficacy. Such experiments can be used not only to test the safety and efficacy of the vaccine in general terms, it can also be used to determine the effective dose.

In general terms, the vaccine is assembled by combining the recombinant virus in a suitable medium or vehicle in

accordance with its intended route of administration. The ingredients are compounded into a medicament in accordance with generally accepted procedures for the preparation of pharmaceutical preparations, as described in standard textbooks on the subject. See, for example, *Pharmaceutical Preformulation and Formulation A Practical Guide* from Candidate Drug Selection to Commercial Dosage Form, M Gibson ed., Informa Health Care 2009, *Pharmaceutical Manufacturing Handbook Production and Processes*, S C Gad ed., Wiley-Interscience 2008, and the latest edition of *Remington's Pharmaceutical Sciences*, Maack Publishing Co, Easton Pa.

Steps in the compounding or formulating of the medicament depend in part on the intended use and mode of administration. Typically, the vaccine will be administered intramuscularly, subcutaneously, or orally. It can be prepared for commercial distribution with any of the following procedures in any effective combination: sterilizing, mixing with appropriate non-toxic and non-interfering excipients, buffers and other carriers, lyophilizing or freezing, dividing into dose units, and enclosing in a delivery device. The medicament will typically be packaged in a suitable container accompanied by or associated with written information about its intended use, such as prophylaxis or treatment of hemorrhagic fever.

A suitable agent as the active ingredient is a modified virus according to this invention as a live virus type vaccine. Alternatively, after replicating in culture, the virus can be inactivated with UV irradiation or chemical means, and the viral particles used with a suitable adjuvant. In essence, attenuation of the vOTU could be used as a safeguard to prevent dangerous live wild type CCHFV from escaping physical attenuation methods for making CCHFV vaccines. The physical attenuation would prevent possible reversion of the virus.

For the purpose of prophylaxis against viral infection, if the subject is adequately primed (such as by previous immunization or infection with the target virus), a single administration of the composition may be sufficient to raise a protective immune response. Multiple administrations are more typical in an immunologically naive host. Desirable outcomes include induction or enhancement of a specific antibody response measured by a suitable test, such as enzyme-linked immunosorbent assay (ELISA) using viral antigens, or a virus neutralization assay.

For purposes of treatment or eradication of an ongoing infectious disease, multiple administrations of the antigen-adjuvant composition (at least 2 or 4, for example, on a biweekly schedule) may be helpful. Here, the objective may be not just to elicit specific antibody, but also to elicit a specific T-lymphocyte response (measured in an ELISPOT™ or proliferation assay), or a cytotoxic T cell response (measurable, for example, in a cytotoxicity assay). Clinical benefit would be manifest as a reduction in the titer of virus or infectious particles in blood or in a tissue biopsy, or a limitation in the progression of necrosis, pain, wasting, or other signs of the disease.

Ultimate choice of the treatment protocol, dose, and monitoring is the responsibility of the managing clinician. Other Genetic Alterations and Other Viruses

CCHF virus and the particular mutations I13R/P77D are used throughout the disclosure for purposes of illustration, and not to limit the practice of the invention.

A person practicing the invention may, as an alternative, change I13 and/or P77 to another amino acid, and/or change other residues in the vOTU protein—so long as the resultant

virus has decreased deubiquitinating activity and/or decreased deISGylating activity, and is still able to replicate in a suitable host cell.

vOTU variants with reduced enzyme activity can be generated by site-directed mutagenesis to introduce a known change into the primary structure if the wild type virus or another variant. The altered virus is then assayed for activity—namely (and in any combination), deubiquitinating activity, deISGylating activity, vOTU protease activity, ability to replicate, and/or ability to suppress cytokines such as interferon β . Thus, another amino acid can be substituted at positions I13 and/or P77, and/or at positions nearby in the tertiary structure. Possible changes include substitutions of one codon for another, and deletions or additions to the encoded amino acid sequence in any combination. Preferred changes will typically retain the tertiary structure of the wild-type virus. For the influence of vOTU structure on enzyme activity, see Capodagli, Pegan et al., *J Virol.* 2013; 87(7):3815-27.

vOTU variants with reduced enzyme activity can also be generated by introducing random mutations into the virus, screening colonies with a functional assay, and selecting colonies with the desired level of enzymatic activity. The particular mutation in the selected virus can then be characterized as to what changes have been made to the viral genome.

Because the genomes of nairoviruses are highly conserved, the invention can also be practiced with other strains of CCHF virus and with other nairoviruses. Possible wild-type nairoviruses that can be modified according to this invention are referred to in various places in this disclosure. Included are the following:

Nairobi Sheep Disease (NSDV; Africa) I Ganjam (Indian variant) is a fatal sheep and goat disease that particularly hinders livestock transport in Africa

Dugbe virus causes mild flu-like symptoms in humans, goats, and sheep. It is present in various parts of Africa and Asia, such as Hazara, Kupe, Dera Ghazi Khan, Hughes, Qalyub, Sakhalin, and Thiafora.

FIGS. 13 and 14 show that quite a number of CCHF viral strains and other nairoviruses are conserved at amino acid positions 13 and 77. Accordingly, the same genetic alterations should have the same biological effects: reduced deubiquitinating and deISGylating activity, while still allowing the virus to replicate.

Besides site directed and random mutagenesis, vOTU variants with reduced enzyme activity can be obtained by building a hybrid virus in which the wild type glycoprotein (M segment) of a nairovirus is replaced with the M segment of another virus having the desired functionality—such as the CCHF I13R/P77D double mutant.

Directed or random changes to a nairovirus genome, and genetic alterations in nairoviruses other than CCHF virus, can be initially screened and tested for vOTU function using assays for deubiquitinating activity and/or deISGylating activity. By way of illustration, a suitable assay for deubiquitination and deISGylation activity can be run as follows. Typically, assays are performed in duplicate in 100 mM NaCl, 50 mM HEPES pH 7.5, 0.01 mg/mL bovine serum albumin (BSA), and 5 mM DTT. A suitable microtiter plate is Corning Costar™ half-volume black 96-well plate with a reaction volume of 50 μ L. The reactions are observed with a matching plate reader, such as an Infinite™ M1000 series reader (Tecan, Inc.). The reaction is followed using ubiquitin

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or other vOTU substrate conjugated to a fluorescent tag, such as 7-amino-4-methylcoumarin (AMC). AMC becomes fluorescent (excitation λ , 360 nm; emission, 460 nm) upon decoupling from the ubiquitin or ISG15.

Suitable substrate conjugates are Ub-AMC, human ISG15-AMC (hISG15-AMC), (Boston Biochem, MA) and ZRLRGG-AMC (SEQ ID NO:22) (Bachem). ZRLRGG (SEQ ID NO:23) is a hexapeptide homologous the carboxy terminal of ubiquitin. Release of AMC is monitored by combining the substrate with wild type (WT) or mutant CCHF vOTU. The extinction coefficients for all three fluorescent substrates can be determined by adding excess vOTU to various concentrations of each substrate and allowing the reactions to run to completion. The resulting maximum fluorescence values are plotted to determine the slope and consequently each substrate's extinction coefficient. Suitable substrate concentrations to measure turnover rates in this assay are of the order of 1 μ M hISG15-AMC with 20 nM vOTU; 1 μ M hUb-AMC with 4 nM vOTU, and 50 μ M ZRLRGG-AMC (SEQ ID NO:22) with 4 μ M vOTU from either wild type or genetically altered virus.

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Advantages

In summary, this invention provides a new technology to produce replicating viral particles suitable for use in a vaccine. Advantages include the following:

CCHF virus with selective mutations can now be produced in human cell lines, avoiding xenogeneic antigen contaminants from animal tissue.

Proven structurally biology-guided mutations of viral ovarian tumor domain proteases ablate deubiquitinating and deISGylating activity.

The recombinant system methodology of this invention can be used to recombinantly generate any nairovirus, or CCHF virus strain. because of the homology.

The method of genetic modification through ablation of deubiquitinating and deISGylating activity can be used in conjunction with physical attenuation methods to ensure a greater level of public safety when administering the vaccine.

SEQUENCES

Reverse genetics system generated Crimean-Congo hemorrhagic fever virus's L-Protein amino acid sequence

(SEQ. ID NO: 1)

MDFLRSLDWTQVRAGQYVSNPRFNISDYFEIVRQPGDGNCFYHSIAELTMPNKTDSYHYIKRLTESAARKYYQEEDEARLVGL
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LSLMDRVIAVDQLTSSSSDELQDYEDLALALTSAEESNRSSLDEVTLSEKKQAEILRQKASQLSKLVNKSQNIPTRVGRVLDGM
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KFEDFLDRTQLHPEFRDLTPDFSLTQKVHFKRNQIPSVENVQISIDATLPESVEAVPVTERRKMPPLPETPLSEVHSIERIMENF
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TTSEKMLLSWLSEDIKSSRCGECLSNIKKAVDETANLSEKIELLAYNLQLTNHCSNCHPNGVNIISNTSNVCKRCPKIEVVSHCE
NKGFEDESNECLTDLRLVRLTLPGKTEKERRVKNVEYLKLMMSMSGIDCIKYPTQLITHGRVSAKHNDGNLKDSDDDQRL
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- continued

LDCGSKEDCKLTLLDLSVSKLLVLCLYQKDDEELINSSSLKLGFLVKYVVTLFTSNGEPFSLSLNDGGDLDDLHKTDEKLLHQ
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Reverse genetics system generated Crimean-Congo hemorrhagic fever virus's M-Protein amino acid sequence

(SEQ. ID NO: 2)

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-continued

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 THIG

Reverse genetics system generated Crimean-Congo hemorrhagic fever virus's S-Protein
 amino acid sequence

(SEQ. ID NO: 3)

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 ETNNPCAktivKLFVQKTGFNIQDMDIVASEHLLHQSLVGKQSPFQNAYNVKGNATSANII

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For all purposes in the United States of America, each and every publication and patent document cited herein is incorporated herein by reference as if each such publication or document was specifically and individually indicated to be incorporated herein by reference.

While the invention has been described with reference to the specific embodiments, changes can be made and equivalents can be substituted to adapt to a particular context or intended use, thereby achieving benefits of the invention without departing from the scope of what is claimed.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 23

<210> SEQ ID NO 1

<211> LENGTH: 3945

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic Nairovirus Crimean-Congo hemorrhagic
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 mutation reducing deubiquinating and deISGylating activities, RNA
 polymerase and viral ovarian tumor domain protease (vOTU)

<220> FEATURE:

<221> NAME/KEY: DOMAIN

<222> LOCATION: (1)...(169)

<223> OTHER INFORMATION: viral ovarian tumor domain protease (vOTU)

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 20 25 30

Arg Gln Pro Gly Asp Gly Asn Cys Phe Tyr His Ser Ile Ala Glu Leu
 35 40 45

Thr Met Pro Asn Lys Thr Asp His Ser Tyr His Tyr Ile Lys Arg Leu
 50 55 60

Thr Glu Ser Ala Ala Arg Lys Tyr Tyr Gln Glu Glu Asp Glu Ala Arg
 65 70 75 80

Leu Val Gly Leu Ser Leu Glu Asp Tyr Leu Lys Arg Met Leu Ser Asp
 85 90 95

Asn Glu Trp Gly Ser Thr Leu Glu Ala Ser Met Leu Ala Lys Glu Met
 100 105 110

Gly Ile Thr Ile Ile Ile Trp Thr Val Ala Ala Ser Asp Glu Val Glu
 115 120 125

Ala Gly Ile Lys Phe Gly Asp Gly Asp Val Phe Thr Ala Val Asn Leu
 130 135 140

Leu His Ser Gly Gln Thr His Phe Asp Ala Leu Arg Ile Leu Pro Gln

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Ala Val Asp Gln Leu Thr Ser Ser Ser Ser Asp Glu Leu Gln Asp Tyr			
	180	185	190
Glu Asp Leu Ala Leu Ala Leu Thr Ser Ala Glu Glu Ser Asn Arg Arg			
	195	200	205
Ser Ser Leu Asp Glu Val Thr Leu Ser Lys Lys Gln Ala Glu Ile Leu			
	210	215	220
Arg Gln Lys Ala Ser Gln Leu Ser Lys Leu Val Asn Lys Ser Gln Asn			
	225	230	235
Ile Pro Thr Arg Val Gly Arg Val Leu Asp Cys Met Phe Asn Cys Lys			
	245	250	255
Leu Cys Val Glu Ile Ser Ala Asp Thr Leu Ile Leu Arg Pro Glu Ser			
	260	265	270
Lys Glu Lys Ile Gly Glu Ile Met Ser Leu Arg Gln Leu Gly His Lys			
	275	280	285
Leu Leu Thr Arg Asp Lys Gln Ile Lys Gln Glu Phe Ser Arg Met Lys			
	290	295	300
Leu Tyr Val Thr Lys Asp Leu Leu Asp His Leu Asp Val Gly Gly Leu			
	305	310	315
Leu Arg Ala Ala Phe Pro Gly Thr Gly Ile Glu Arg His Met Gln Leu			
	325	330	335
Leu His Ser Glu Met Ile Leu Asp Ile Cys Thr Val Ser Leu Gly Val			
	340	345	350
Met Leu Ser Thr Phe Leu Tyr Gly Ser Asn Asn Lys Asn Lys Lys Lys			
	355	360	365
Phe Ile Thr Asn Cys Leu Leu Ser Thr Ala Leu Ser Gly Lys Lys Val			
	370	375	380
Tyr Lys Val Leu Gly Asn Leu Gly Asn Glu Leu Leu Tyr Lys Ala Pro			
	385	390	395
Arg Lys Ala Leu Ala Thr Val Cys Ser Ala Leu Phe Gly Lys Gln Ile			
	405	410	415
Asn Lys Leu Gln Asn Cys Phe Arg Thr Ile Ser Pro Val Ser Leu Leu			
	420	425	430
Ala Leu Arg Asn Leu Asp Phe Asp Cys Leu Ser Val Gln Asp Tyr Asn			
	435	440	445
Gly Met Ile Glu Asn Met Ser Lys Leu Asp Asn Thr Asp Val Glu Phe			
	450	455	460
Asn His Arg Glu Ile Ala Asp Leu Asn Gln Leu Thr Ser Arg Leu Ile			
	465	470	475
Thr Leu Arg Lys Glu Lys Asp Thr Asp Leu Leu Lys Gln Trp Phe Pro			
	485	490	495
Glu Ser Asp Leu Thr Arg Arg Ser Ile Arg Asn Ala Ala Asn Ala Glu			
	500	505	510
Glu Phe Val Ile Ser Glu Phe Phe Lys Lys Lys Asp Ile Met Lys Phe			
	515	520	525
Ile Ser Thr Ser Gly Arg Ala Met Ser Ala Gly Lys Ile Gly Asn Val			
	530	535	540
Leu Ser Tyr Ala His Asn Leu Tyr Leu Ser Lys Ser Ser Leu Asn Met			
	545	550	555
Thr Ser Glu Asp Ile Ser Gln Leu Leu Ile Glu Ile Lys Arg Leu Tyr			
	565	570	575

Ala 580	Leu	Gln	Glu	Asp	Ser	Glu	Val	Glu 585	Pro	Ile	Ala	Ile	Ile 590	Cys	Asp
Gly 595	Ile	Glu	Ser	Asn	Met	Lys	Gln 600	Leu	Phe	Ala	Ile	Leu 605	Pro	Pro	Asp
Cys 610	Ala	Arg	Glu	Cys	Glu	Val 615	Leu	Phe	Asp	Asp	Ile 620	Arg	Asn	Ser	Pro
Thr 625	His	Ser	Thr	Ala	Trp 630	Lys	His	Ala	Leu	Arg 635	Leu	Lys	Gly	Thr	Ala 640
Tyr	Glu	Gly	Leu	Phe 645	Ala	Asn	Cys	Tyr	Gly 650	Trp	Gln	Tyr	Ile	Pro 655	Glu
Asp	Ile	Lys	Pro 660	Ser	Leu	Thr	Met 665	Leu	Ile	Gln	Thr	Leu	Phe 670	Pro	Asp
Lys	Phe 675	Glu	Asp	Phe	Leu	Asp	Arg 680	Thr	Gln	Leu	His	Pro 685	Glu	Phe	Arg
Asp 690	Leu	Thr	Pro	Asp	Phe 695	Ser	Leu	Thr	Gln	Lys	Val 700	His	Phe	Lys	Arg
Asn 705	Gln	Ile	Pro	Ser	Val 710	Glu	Asn	Val	Gln	Ile 715	Ser	Ile	Asp	Ala	Thr 720
Leu	Pro	Glu	Ser	Val 725	Glu	Ala	Val	Pro	Val 730	Thr	Glu	Arg	Lys	Met 735	Phe
Pro	Leu	Pro	Glu	Thr 740	Pro	Leu	Ser	Glu 745	Val	His	Ser	Ile 750	Glu	Arg	Ile
Met	Glu	Asn 755	Phe	Thr	Arg	Leu	Met 760	His	Gly	Gly	Arg	Leu 765	Ser	Thr	Lys
Lys 770	Arg	Asp	Gly	Asp	Pro	Ala 775	Glu	Gln	Gly	Asn	Gln 780	Gln	Ser	Ile	Thr
Glu 785	His	Glu	Ser	Ser	Ser 790	Ile	Ser	Ala	Phe	Lys 795	Asp	Tyr	Gly	Glu	Arg 800
Gly	Ile	Val	Glu	Glu 805	Asn	His	Met	Lys	Phe	Ser	Gly	Glu	Asp	Gln	Leu 815
Glu	Thr	Arg	Gln	Leu	Leu 820	Leu	Val	Glu 825	Val	Gly	Phe	Gln	Thr	Asp	Ile 830
Asp	Gly	Lys 835	Ile	Arg	Thr	Asp	His 840	Lys	Lys	Trp	Lys	Asp 845	Ile	Leu	Lys
Leu 850	Leu	Glu	Leu	Leu	Gly 855	Ile	Lys	Cys	Ser	Phe	Ile	Ala	Cys	Ala	Asp
Cys 865	Ser	Ser	Thr	Pro	Pro 870	Asp	Arg	Trp	Trp	Ile 875	Thr	Glu	Asp	Arg	Val 880
Arg	Val	Leu	Lys	Asn 885	Ser	Val	Ser	Phe	Leu	Phe	Asn	Lys	Leu	Ser	Arg 895
Asn	Ser	Pro	Thr	Glu	Val 900	Thr	Asp	Ile 905	Val	Val	Gly	Ala	Ile	Ser	Thr 910
Gln	Lys	Val	Arg	Ser	Tyr 915	Leu	Lys	Ala 920	Gly	Thr	Ala	Thr 925	Lys	Thr	Pro
Val 930	Ser	Thr	Lys	Asp	Val 935	Leu	Glu	Thr	Trp	Glu	Lys	Met 940	Lys	Glu	His
Ile 945	Leu	Asn	Arg	Pro	Thr 950	Gly	Leu	Thr	Leu	Pro 955	Thr	Ser	Leu	Glu	Gln 960
Ala	Met	Arg	Lys	Gly 965	Leu	Val	Glu	Gly	Val	Val	Ile	Ser	Lys	Glu	Gly 975
Ser	Glu	Ser	Cys	Ile	Asn 980	Met	Leu	Lys 985	Glu	Asn	Leu	Asp	Arg	Ile	Thr 990

Asp	Glu	Phe	Glu	Arg	Thr	Lys	Phe	Lys	His	Glu	Leu	Thr	Gln	Asn	Ile
	995						1000					1005			
Thr	Thr	Ser	Glu	Lys	Met	Leu	Leu	Ser	Trp	Leu	Ser	Glu	Asp	Ile	Lys
	1010					1015					1020				
Ser	Ser	Arg	Cys	Gly	Glu	Cys	Leu	Ser	Asn	Ile	Lys	Lys	Ala	Val	Asp
	1025				1030					1035					1040
Glu	Thr	Ala	Asn	Leu	Ser	Glu	Lys	Ile	Glu	Leu	Leu	Ala	Tyr	Asn	Leu
				1045					1050					1055	
Gln	Leu	Thr	Asn	His	Cys	Ser	Asn	Cys	His	Pro	Asn	Gly	Val	Asn	Ile
			1060					1065					1070		
Ser	Asn	Thr	Ser	Asn	Val	Cys	Lys	Arg	Cys	Pro	Lys	Ile	Glu	Val	Val
	1075						1080					1085			
Ser	His	Cys	Glu	Asn	Lys	Gly	Phe	Glu	Asp	Ser	Asn	Glu	Cys	Leu	Thr
	1090					1095					1100				
Asp	Leu	Asp	Arg	Leu	Val	Arg	Leu	Thr	Leu	Pro	Gly	Lys	Thr	Glu	Lys
	1105				1110					1115					1120
Glu	Arg	Arg	Val	Lys	Arg	Asn	Val	Glu	Tyr	Leu	Ile	Lys	Leu	Met	Met
				1125					1130					1135	
Ser	Met	Ser	Gly	Ile	Asp	Cys	Ile	Lys	Tyr	Pro	Thr	Gly	Gln	Leu	Ile
			1140					1145					1150		
Thr	His	Gly	Arg	Val	Ser	Ala	Lys	His	Asn	Asp	Gly	Asn	Leu	Lys	Asp
	1155						1160					1165			
Arg	Ser	Asp	Asp	Asp	Gln	Arg	Leu	Ala	Glu	Lys	Ile	Asp	Thr	Val	Arg
	1170					1175					1180				
Lys	Glu	Leu	Ser	Glu	Ser	Lys	Leu	Lys	Asp	Tyr	Ser	Thr	Tyr	Ala	Arg
	1185				1190				1195						1200
Gly	Val	Ile	Ser	Asn	Ser	Leu	Lys	Asn	Leu	Ser	Arg	Gln	Gly	Lys	Ser
				1205					1210					1215	
Lys	Cys	Ser	Val	Pro	Arg	Ser	Trp	Leu	Glu	Lys	Val	Leu	Phe	Asp	Leu
			1220					1225					1230		
Lys	Val	Pro	Thr	Lys	Asp	Glu	Glu	Val	Leu	Ile	Asn	Ile	Arg	Asn	Ser
	1235						1240					1245			
Leu	Lys	Ala	Arg	Ser	Glu	Phe	Val	Arg	Asn	Asn	Asp	Lys	Leu	Leu	Ile
	1250					1255					1260				
Arg	Ser	Lys	Glu	Glu	Leu	Lys	Lys	Cys	Phe	Asp	Val	Gln	Ser	Phe	Lys
	1265				1270				1275						1280
Leu	Lys	Lys	Asn	Lys	Gln	Pro	Val	Pro	Phe	Gln	Val	Asp	Cys	Ile	Leu
				1285					1290					1295	
Phe	Lys	Glu	Val	Ala	Ala	Glu	Cys	Met	Lys	Arg	Tyr	Ile	Gly	Thr	Pro
			1300					1305					1310		
Tyr	Glu	Gly	Ile	Val	Asp	Thr	Leu	Val	Ser	Leu	Ile	Asn	Val	Leu	Thr
	1315						1320					1325			
Arg	Phe	Thr	Trp	Phe	Gln	Glu	Val	Val	Leu	Tyr	Gly	Lys	Ile	Cys	Glu
	1330					1335									

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1410	1415	1420
Gln Tyr Arg Cys Leu Glu Val Ile Asn Ser Val Ser Glu Lys Thr Leu		
1425	1430	1435 1440
Gln Asp Ile Glu Asn His Ser Met Thr Leu Leu Glu Asp Ser Phe Arg		
	1445	1450 1455
Glu Ile Thr Phe Ala Leu Glu Gly Arg Phe Glu Glu Ser Tyr Lys Ile		
	1460	1465 1470
Arg Thr Ser Arg Cys Arg Ala Ser Gly Asn Phe Leu Asn Arg Ser Ser		
	1475	1480 1485
Arg Asp His Phe Ile Ser Val Val Ser Gly Leu Asn Leu Val Tyr Gly		
	1490	1495 1500
Phe Leu Ile Lys Asp Asn Leu Leu Ala Asn Ser Gln Gln Gln Asn Lys		
1505	1510	1515 1520
Gln Leu Gln Met Leu Arg Phe Gly Met Leu Ala Gly Leu Ser Arg Leu		
	1525	1530 1535
Val Cys Pro Asn Glu Leu Gly Lys Lys Phe Ser Thr Ser Cys Arg Arg		
	1540	1545 1550
Ile Glu Asp Asn Ile Ala Arg Leu Tyr Leu Gln Thr Ser Ile Tyr Cys		
	1555	1560 1565
Ser Val Arg Asp Val Glu Asp Asn Val Lys His Trp Lys Gln Arg Asp		
	1570	1575 1580
Leu Cys Pro Glu Val Thr Ile Pro Cys Phe Thr Val Tyr Gly Thr Phe		
1585	1590	1595 1600
Val Asn Ser Asp Arg Gln Leu Ile Phe Asp Ile Tyr Asn Val His Ile		
	1605	1610 1615
Tyr Asn Lys Glu Met Asp Asn Phe Asp Glu Gly Cys Ile Ser Val Leu		
	1620	1625 1630
Glu Glu Thr Ala Glu Arg His Met Leu Trp Glu Leu Asp Leu Met Asn		
	1635	1640 1645
Ser Leu Cys Ser Asp Glu Lys Lys Asp Thr Arg Thr Ala Arg Leu Leu		
	1650	1655 1660
Leu Gly Cys Pro Asn Val Arg Lys Ala Ala Asn Arg Glu Gly Lys Lys		
1665	1670	1675 1680
Leu Leu Lys Leu Asn Ser Asp Thr Ser Thr Asp Thr Gln Ser Ile Ala		
	1685	1690 1695
Ser Glu Val Ser Asp Arg Arg Ser Tyr Ser Ser Ser Lys Ser Arg Ile		
	1700	1705 1710
Arg Ser Ile Phe Gly Arg Tyr Asn Ser Gln Lys Lys Pro Phe Glu Leu		
	1715	1720 1725
Arg Ser Gly Leu Glu Val Phe Asn Asp Pro Phe Asn Asp Tyr Gln Gln		
	1730	1735 1740
Ala Ile Thr Asp Ile Cys Gln Phe Ser Glu Tyr Thr Pro Asn Lys Glu		
1745	1750	1755 1760
Ser Ile Leu Lys Asp Cys Leu Gln Ile Ile Arg Lys Asn Pro Ser His		
	1765	1770 1775
Thr Met Gly Ser Phe Glu Leu Ile Gln Ala Ile Ser Glu Phe Gly Met		
	1780	1785 1790
Ser Lys Phe Pro Pro Glu Asn Ile Asp Lys Ala Arg Arg Asp Pro Lys		
	1795	1800 1805
Asn Trp Val Ser Ile Ser Glu Val Thr Glu Thr Thr Ser Ile Val Ala		
	1810	1815 1820
Ser Pro Arg Thr His Met Met Leu Lys Asp Cys Phe Lys Ile Ile Leu		
1825	1830	1835 1840

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Gly Thr Glu Asn Lys Lys Ile Val Lys Met Leu Arg Gly Lys Leu Lys	1845	1850	1855
Lys Leu Gly Ala Ile Ser Thr Asn Ile Glu Ile Gly Lys Arg Asp Cys	1860	1865	1870
Leu Asp Leu Leu Ser Thr Val Asp Gly Leu Thr Asp Gln Gln Lys Glu	1875	1880	1885
Asn Ile Val Asn Gly Ile Phe Glu Pro Ser Lys Leu Ser Phe Tyr His	1890	1895	1900
Trp Lys Glu Leu Val Lys Lys Asn Ile Asp Glu Val Leu Leu Thr Glu	1905	1910	1915
Asp Gly Asn Leu Ile Phe Cys Trp Leu Lys Thr Ile Ser Ser Ser Val	1925	1930	1935
Lys Gly Ser Leu Lys Lys Arg Leu Lys Phe Met Asn Ile His Ser Pro	1940	1945	1950
Glu Leu Met Pro Glu Asn Cys Leu Phe Ser Ser Glu Glu Phe Asn Glu	1955	1960	1965
Leu Ile Lys Leu Lys Lys Leu Leu Leu Asn Glu Gln Gln Asp Glu Gln	1970	1975	1980
Glu Leu Lys Gln Asp Leu Leu Ile Ser Ser Trp Ile Lys Cys Ile Thr	1985	1990	1995
Ala Cys Lys Asp Phe Ala Ser Ile Asn Asp Lys Ile Gln Lys Phe Ile	2005	2010	2015
Tyr His Leu Ser Glu Glu Leu Tyr Asp Ile Arg Leu Gln His Leu Glu	2020	2025	2030
Leu Ser Lys Leu Lys Gln Glu His Pro Ser Val Ser Phe Thr Lys Glu	2035	2040	2045
Glu Val Leu Ile Lys Arg Leu Glu Lys Asn Phe Leu Lys Gln His Asn	2050	2055	2060
Leu Glu Ile Met Glu Thr Val Asn Leu Val Phe Phe Ala Ala Leu Ser	2065	2070	2075
Ala Pro Trp Cys Leu His Tyr Lys Ala Leu Glu Ser Tyr Leu Val Arg	2085	2090	2095
His Pro Glu Ile Leu Asp Cys Gly Ser Lys Glu Asp Cys Lys Leu Thr	2100	2105	2110
Leu Leu Asp Leu Ser Val Ser Lys Leu Leu Val Cys Leu Tyr Gln Lys	2115	2120	2125
Asp Asp Glu Glu Leu Ile Asn Ser Ser Ser Leu Lys Leu Gly Phe Leu	2130	2135	2140
Val Lys Tyr Val Val Thr Leu Phe Thr Ser Asn Gly Glu Pro Phe Ser	2145	2150	2155
Leu Ser Leu Asn Asp Gly Gly Leu Asp Leu Asp Leu His Lys Thr Thr	2165	2170	2175
Asp Glu Lys Leu Leu His Gln Thr Lys Ile Val Phe Ala Lys Ile Gly	2180	2185	2190
Leu Ser Gly Asn Ser Tyr Asp Phe Ile Trp Thr Thr Gln Met Ile Ala	2195	2200	2205
Asn Ser Asn Phe Asn Val Cys Lys Arg Leu Thr Gly Arg Ser Thr Gly	2210	2215	2220
Glu Arg Leu Pro Arg Ser Val Arg Ser Lys Val Ile Tyr Glu Met Val	2225	2230	2235
Lys Leu Val Gly Glu Thr Gly Met Ala Ile Leu Gln Gln Leu Ala Phe	2245	2250	2255

Ala	Gln	Ala	Leu	Asn	Tyr	Glu	His	Arg	Phe	Tyr	Ala	Val	Leu	Ala	Pro
			2260					2265					2270		
Lys	Ala	Gln	Leu	Gly	Gly	Ala	Arg	Asp	Leu	Leu	Val	Gln	Glu	Thr	Gly
	2275						2280					2285			
Thr	Lys	Val	Met	His	Ala	Thr	Thr	Glu	Met	Phe	Ser	Arg	Asn	Leu	Leu
	2290					2295					2300				
Lys	Thr	Thr	Ser	Asp	Asp	Gly	Leu	Thr	Asn	Pro	His	Leu	Lys	Glu	Thr
2305					2310					2315					2320
Ile	Leu	Asn	Val	Gly	Leu	Asp	Cys	Leu	Ala	Asn	Met	Arg	Asn	Leu	Asp
			2325						2330					2335	
Gly	Lys	Pro	Ile	Ser	Glu	Gly	Ser	Asn	Leu	Val	Asn	Phe	Tyr	Lys	Val
		2340						2345					2350		
Ile	Cys	Ile	Ser	Gly	Asp	Asn	Thr	Lys	Trp	Gly	Pro	Ile	His	Cys	Cys
	2355					2360						2365			
Ser	Phe	Phe	Ser	Gly	Met	Met	Gln	Gln	Val	Leu	Lys	Asn	Val	Pro	Asp
	2370					2375					2380				
Trp	Cys	Ser	Phe	Tyr	Lys	Leu	Thr	Phe	Ile	Lys	Asn	Leu	Cys	Arg	Gln
2385					2390					2395					2400
Val	Glu	Ile	Pro	Ala	Gly	Ser	Ile	Lys	Lys	Ile	Leu	Asn	Val	Leu	Arg
			2405						2410					2415	
Tyr	Arg	Leu	Cys	Ser	Lys	Gly	Gly	Val	Glu	Gln	His	Ser	Glu	Glu	Asp
		2420						2425					2430		
Leu	Arg	Arg	Leu	Leu	Thr	Asp	Asn	Leu	Asp	Ser	Trp	Asp	Gly	Asn	Asp
	2435						2440					2445			
Thr	Val	Lys	Phe	Leu	Val	Thr	Thr	Tyr	Ile	Ser	Lys	Gly	Leu	Met	Ala
	2450					2455					2460				
Leu	Asn	Ser	Tyr	Asn	His	Met	Gly	Gln	Gly	Ile	His	His	Ala	Thr	Ser
2465					2470					2475					2480
Ser	Val	Leu	Thr	Ser	Leu	Ala	Ala	Val	Leu	Phe	Glu	Glu	Leu	Ala	Ile
			2485					2490						2495	
Phe	Tyr	Leu	Lys	Arg	Ser	Leu	Pro	Gln	Thr	Thr	Val	His	Val	Glu	His
		2500						2505					2510		
Ala	Gly	Ser	Ser	Asp	Asp	Tyr	Ala	Lys	Cys	Ile	Val	Val	Thr	Gly	Ile
	2515						2520					2525			
Leu	Ser	Lys	Glu	Leu	Tyr	Ser	Gln	Tyr	Asp	Glu	Thr	Phe	Trp	Lys	His
	2530					2535					2540				
Ala	Cys	Arg	Leu	Lys	Asn	Phe	Thr	Ala	Ala	Val	Gln	Arg	Cys	Cys	Gln
2545					2550					2555					2560
Met	Lys	Asp	Ser	Ala	Lys	Thr	Leu	Val	Ser	Asp	Cys	Phe	Leu	Glu	Phe
			2565					2570					2575		
Tyr	Ser	Glu	Phe	Met	Met	Gly	Tyr	Arg	Val	Thr	Pro	Ala	Val	Ile	Lys
		2580						2585					2590		
Phe	Met	Phe	Thr	Gly	Leu	Ile	Asn	Ser	Ser	Val	Thr	Ser	Pro	Gln	Ser
	2595														

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2675	2680	2685
Arg Ala Ala Gln Thr Leu Gln Met Asn Ser Val Ser Ile Gln Ser Ser		
2690	2695	2700
Ser Leu Thr Thr Leu Asp Ser Leu Gly Arg Ser Arg Thr Ser Ser Thr		
2705	2710	2715 2720
Ala Glu Asp Ser Ser Ser Val Ser Asp Thr Thr Ala Ala Ser His Asp		
	2725	2730 2735
Ser Gly Ser Ser Ser Ser Ser Phe Ser Phe Glu Leu Asn Arg Pro Leu		
	2740	2745 2750
Ser Glu Thr Glu Leu Gln Phe Ile Lys Ala Leu Ser Ser Leu Lys Ser		
	2755	2760 2765
Thr Gln Ala Cys Glu Val Ile Gln Asn Arg Ile Thr Gly Leu Tyr Cys		
	2770	2775 2780
Asn Ser Asn Glu Gly Pro Leu Asp Arg His Asn Val Ile Tyr Ser Ser		
2785	2790	2795 2800
Arg Met Ala Asp Ser Cys Asp Trp Leu Lys Asp Gly Lys Arg Arg Gly		
	2805	2810 2815
Asn Leu Glu Leu Ala Asn Arg Ile Gln Ser Val Leu Cys Ile Leu Ile		
	2820	2825 2830
Ala Gly Tyr Tyr Arg Ser Phe Gly Gly Glu Gly Thr Glu Lys Gln Val		
	2835	2840 2845
Lys Ala Ser Leu Asn Arg Asp Asp Asn Lys Ile Ile Glu Asp Pro Met		
	2850	2855 2860
Ile Gln Leu Ile Pro Glu Lys Leu Arg Arg Glu Leu Glu Arg Leu Gly		
2865	2870	2875 2880
Val Ser Arg Met Glu Val Asp Glu Leu Met Pro Ser Ile Ser Pro Asp		
	2885	2890 2895
Asp Thr Leu Ala Gln Leu Val Ala Lys Lys Leu Ile Ser Leu Asn Val		
	2900	2905 2910
Ser Thr Glu Glu Tyr Ser Ala Glu Val Ser Arg Leu Lys Gln Thr Leu		
	2915	2920 2925
Thr Ala Arg Asn Val Leu His Gly Leu Ala Gly Gly Ile Lys Glu Leu		
	2930	2935 2940
Ser Leu Pro Ile Tyr Thr Ile Phe Met Lys Ser Tyr Phe Phe Lys Asp		
2945	2950	2955 2960
Asn Val Phe Leu Ser Leu Thr Asp Arg Trp Ser Thr Lys His Ser Thr		
	2965	2970 2975
Asn Tyr Arg Asp Ser Cys Gly Lys Gln Leu Lys Gly Arg Ile Ile Thr		
	2980	2985 2990
Lys Tyr Thr His Trp Leu Asp Thr Phe Leu Gly Cys Ser Val Ser Ile		
	2995	3000 3005
Asn Arg His Thr Thr Val Lys Glu Pro Ser Leu Phe Asn Pro Asn Ile		
	3010	3015 3020
Arg Cys Val Asn Leu Ile Thr Phe Glu Asp Gly Leu Arg Glu Leu Ser		
3025	3030	3035 3040
Val Ile Gln Ser His Leu Lys Val Phe Glu Asn Glu Phe Thr Asn Leu		
	3045	3050 3055
Asn Leu Gln Phe Ser Asp Pro Asn Arg Gln Lys Leu Arg Ile Val Glu		
	3060	3065 3070
Ser Arg Pro Ala Glu Ser Glu Leu Glu Ala Asn Arg Ala Val Ile Val		
	3075	3080 3085
Lys Thr Lys Leu Phe Ser Ala Thr Glu Gln Val Arg Leu Ser Asn Asn		
	3090	3095 3100

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Pro Ala Val Val Met Gly Tyr Leu Leu Asp Glu Ser Ala Ile Ser Glu			
3105	3110	3115	3120
Val Lys Pro Thr Lys Val Asp Phe Ser Asn Leu Leu Lys Asp Arg Phe			
	3125	3130	3135
Lys Ile Met Gln Phe Phe Pro Ser Val Phe Thr Leu Ile Lys Met Leu			
	3140	3145	3150
Thr Asp Glu Ser Ser Asp Ser Glu Lys Ser Gly Leu Ser Pro Asp Leu			
	3155	3160	3165
Gln Gln Val Ala Arg Tyr Ser Asn His Leu Thr Leu Leu Ser Arg Met			
	3170	3175	3180
Ile Gln Gln Ala Lys Pro Thr Val Thr Val Phe Tyr Met Leu Lys Gly			
3185	3190	3195	3200
Asn Leu Met Asn Thr Glu Pro Thr Val Ala Glu Leu Val Ser Tyr Gly			
	3205	3210	3215
Ile Lys Glu Gly Arg Phe Phe Arg Leu Ser Asp Thr Gly Val Asp Ala			
	3220	3225	3230
Ser Thr Tyr Ser Val Lys Tyr Trp Lys Ile Leu His Cys Ile Ser Ala			
	3235	3240	3245
Ile Gly Cys Leu Pro Leu Ser Gln Ala Asp Lys Ser Ser Leu Leu Met			
	3250	3255	3260
Ser Phe Leu Asn Trp Arg Val Asn Met Asp Ile Arg Thr Ser Asp Cys			
3265	3270	3275	3280
Pro Leu Ser Ser His Glu Ala Ser Ile Leu Ser Glu Phe Asp Gly Gln			
	3285	3290	3295
Val Ile Ala Asn Ile Leu Ala Ser Glu Leu Ser Ser Val Lys Arg Asp			
	3300	3305	3310
Ser Glu Arg Glu Gly Leu Thr Asp Leu Leu Asp Tyr Leu Asn Ser Pro			
	3315	3320	3325
Thr Glu Leu Leu Lys Lys Lys Pro Tyr Leu Gly Thr Thr Cys Lys Phe			
	3330	3335	3340
Asn Thr Trp Gly Asp Ser Asn Arg Ser Gly Lys Phe Thr Tyr Ser Ser			
3345	3350	3355	3360
Arg Ser Gly Glu Ser Ile Gly Ile Phe Ile Ala Gly Lys Leu His Ile			
	3365	3370	3375
His Leu Ser Ser Glu Ser Val Ala Leu Leu Cys Glu Thr Glu Arg Gln			
	3380	3385	3390
Val Leu Ser Trp Met Ser Lys Arg Arg Thr Glu Val Ile Thr Lys Glu			
	3395	3400	3405
Gln His Gln Leu Phe Leu Ser Leu Leu Pro Gln Ser His Glu Cys Leu			
	3410	3415	3420
Gln Lys His Lys Asp Gly Ser Ala Leu Ser Val Ile Pro Asp Ser Ser			
3425	3430	3435	3440
Asn Pro Arg Leu Leu Lys Phe Val Pro Leu Lys Lys Gly Leu Ala Val			
	3445	3450	3455
Val Lys Ile Lys Lys Gln Ile Leu Thr Val Lys Lys Gln Val Val Phe			
	3460	3465	3470
Asp Ala Glu Ser Glu Pro Arg Leu Gln Trp Gly His Gly Cys Leu Ser			
	3475	3480	3485
Ile Val Tyr Asp Glu Thr Asp Thr Gln Thr Thr Tyr His Glu Asn Leu			
	3490	3495	3500
Leu Lys Val Lys His Leu Val Asp Cys Ser Thr Asp Arg Lys Lys Leu			
3505	3510	3515	3520

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Leu Pro Gln Ser Val Phe Ser Asp Ser Lys Val Val Leu Ser Arg Ile
 3525 3530 3535
 Lys Phe Lys Thr Glu Leu Leu Leu Asn Ser Leu Thr Leu Leu His Cys
 3540 3545 3550
 Phe Leu Lys His Ala Pro Ser Asp Ala Ile Met Glu Val Glu Ser Lys
 3555 3560 3565
 Ser Ser Leu Leu His Lys Tyr Leu Lys Ser Gly Gly Val Arg Gln Arg
 3570 3575 3580
 Asn Thr Glu Val Leu Phe Arg Glu Lys Leu Asn Lys Val Val Ile Lys
 3585 3590 3595 3600
 Asp Asn Leu Glu Gln Gly Val Glu Glu Glu Ile Glu Phe Cys Asn Asn
 3605 3610 3615
 Leu Thr Lys Thr Val Ser Glu Asn Pro Leu Pro Leu Ser Cys Trp Ser
 3620 3625 3630
 Glu Val Gln Asn Tyr Ile Glu Asp Ile Gly Phe Asn Asn Val Leu Val
 3635 3640 3645
 Asn Ile Asp Arg Asn Thr Val Lys Ser Glu Leu Leu Trp Lys Phe Thr
 3650 3655 3660
 Leu Asp Thr Asn Val Ser Thr Thr Ser Thr Ile Lys Asp Val Arg Thr
 3665 3670 3675 3680
 Leu Val Ser Tyr Val Ser Thr Glu Thr Ile Pro Lys Phe Leu Leu Ala
 3685 3690 3695
 Phe Leu Leu Tyr Glu Glu Val Leu Met Asn Leu Ile Asn Gln Cys Lys
 3700 3705 3710
 Ala Val Lys Glu Leu Ile Asn Ser Thr Gly Leu Ser Asp Leu Glu Leu
 3715 3720 3725
 Glu Ser Leu Leu Thr Leu Cys Ala Phe Tyr Phe Gln Ser Glu Cys Ser
 3730 3735 3740
 Lys Arg Asp Gly Pro Arg Cys Ser Phe Ala Ala Leu Leu Ser Leu Ile
 3745 3750 3755 3760
 His Glu Asp Trp Gln Arg Ile Gly Lys Asn Ile Leu Val Arg Ala Asn
 3765 3770 3775
 Asn Glu Leu Gly Asp Val Ser Leu Lys Val Asn Ile Val Leu Val Pro
 3780 3785 3790
 Leu Lys Asp Met Ser Lys Pro Lys Ser Glu Arg Val Val Met Ala Arg
 3795 3800 3805
 Arg Ser Leu Asn His Ala Leu Ser Leu Met Phe Leu Asp Glu Met Ser
 3810 3815 3820
 Leu Pro Glu Leu Lys Ser Leu Ser Val Asn Cys Lys Met Gly Asn Phe
 3825 3830 3835 3840
 Glu Gly Gln Glu Cys Phe Glu Phe Thr Ile Leu Lys Asp Asn Ser Ala
 3845 3850 3855
 Arg Leu Asp Tyr Asn Lys Leu Ile Asp His Cys Val Asp Met Glu Lys
 3860 3865 3870
 Lys Arg Glu Ala Val Arg Ala Val Glu Asp Leu Ile Leu Met Leu Thr
 3875 3880 3885
 Gly Arg Ala Val Lys Pro Ser Ala Val Thr Gln Phe Val His Gly Asp
 3890 3895 3900
 Glu Gln Cys Gln Glu Ile Ser Leu Asp Asp Leu Met Ala Asn Asp
 3905 3910 3915 3920
 Thr Val Thr Asp Phe Pro Asp Arg Glu Ala Glu Ala Leu Lys Thr Gly
 3925 3930 3935
 Asn Leu Gly Phe Asn Trp Asp Ser Asp

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3940
<210> SEQ ID NO 2
<211> LENGTH: 1684
<212> TYPE: PRT
<213> ORGANISM: Nairovirus
<220> FEATURE:
<223> OTHER INFORMATION: Nairovirus Crimean-Congo hemorrhagic fever
      virus (CCHFV) M (Middle) protein, envelope proteins Ge
      and Gn glycoprotein precursor

<400> SEQUENCE: 2

Met His Ile Ser Leu Met Tyr Ala Ile Leu Cys Leu Gln Leu Cys Gly
 1             5             10             15
Leu Gly Glu Thr His Gly Ser His Asn Glu Thr Arg His Asn Lys Thr
 20            25            30
Asp Thr Met Thr Thr Pro Gly Asp Asn Pro Ser Ser Glu Pro Pro Val
 35            40            45
Ser Thr Ala Leu Ser Ile Thr Leu Asp Pro Ser Thr Val Thr Pro Thr
 50            55            60
Thr Pro Ala Ser Gly Leu Glu Gly Ser Gly Glu Val Tyr Thr Ser Pro
 65            70            75            80
Pro Ile Thr Thr Gly Ser Leu Pro Leu Ser Glu Thr Thr Pro Glu Leu
 85            90            95
Pro Val Thr Thr Gly Thr Asp Thr Leu Ser Ala Gly Asp Val Asp Pro
100           105           110
Ser Thr Gln Thr Ala Gly Gly Thr Ser Ala Pro Thr Val Arg Thr Ser
115           120           125
Leu Pro Asn Ser Pro Ser Thr Pro Ser Thr Pro Gln Asp Thr His His
130           135           140
Pro Val Arg Asn Leu Leu Ser Val Thr Ser Pro Gly Pro Asp Glu Thr
145           150           155           160
Ser Thr Pro Ser Gly Thr Gly Lys Glu Ser Ser Ala Thr Ser Ser Pro
165           170           175
His Pro Val Ser Asn Arg Pro Pro Thr Pro Pro Ala Thr Ala Gln Gly
180           185           190
Pro Thr Glu Asn Asp Ser His Asn Ala Thr Glu His Pro Glu Ser Leu
195           200           205
Thr Gln Ser Ala Thr Pro Gly Leu Met Thr Ser Pro Thr Gln Ile Val
210           215           220
His Pro Gln Ser Ala Thr Pro Ile Thr Val Gln Asp Thr His Pro Ser
225           230           235           240
Pro Thr Asn Arg Ser Lys Arg Asn Leu Lys Met Glu Ile Ile Leu Thr
245           250           255
Leu Ser Gln Gly Leu Lys Lys Tyr Tyr Gly Lys Ile Leu Arg Leu Leu
260           265           270
Gln Leu Thr Leu Glu Glu Asp Thr Glu Gly Leu Leu Glu Trp Cys Lys
275           280           285
Arg Asn Leu Gly Leu Asp Cys Asp Asp Thr Phe Phe Gln Lys Arg Ile
290           295           300
Glu Glu Phe Phe Ile Thr Gly Glu Gly His Phe Asn Glu Val Leu Gln
305           310           315           320
Phe Arg Thr Pro Gly Thr Leu Ser Thr Thr Glu Ser Thr Pro Ala Gly
325           330           335
Leu Pro Thr Ala Glu Pro Phe Lys Ser Tyr Phe Ala Lys Gly Phe Leu
340           345           350

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Ser 355	Ile	Asp	Ser	Gly	Tyr	Tyr	Ser 360	Ala	Lys	Cys	Tyr	Ser 365	Gly	Thr	Ser
Asn 370	Ser	Gly	Leu	Gln	Leu	Ile 375	Asn	Ile	Thr	Arg	His 380	Ser	Thr	Arg	Ile
Val 385	Asp	Thr	Pro	Gly	Pro 390	Lys	Ile	Thr	Asn	Leu 395	Lys	Thr	Ile	Asn	Cys 400
Ile	Asn	Leu	Lys	Ala 405	Ser	Ile	Phe	Lys	Glu 410	His	Arg	Glu	Val	Glu 415	Ile
Asn	Val	Leu	Leu	Pro	Gln	Val	Ala	Val 425	Asn	Leu	Ser	Asn	Cys 430	His	Val
Val	Ile	Lys	Ser	His	Val	Cys	Asp 440	Tyr	Ser	Leu	Asp	Ile 445	Asp	Gly	Ala
Val	Arg	Leu	Pro	His	Ile	Tyr 455	His	Glu	Gly	Val	Phe	Ile 460	Pro	Gly	Thr
Tyr 465	Lys	Ile	Val	Ile	Asp 470	Lys	Lys	Asn	Lys	Leu 475	Asn	Asp	Arg	Cys	Thr 480
Leu	Phe	Thr	Asp	Cys 485	Val	Ile	Lys	Gly	Arg	Glu 490	Val	Arg	Lys	Gly 495	Gln
Ser	Val	Leu	Arg	Gln	Tyr	Lys	Thr	Glu 505	Ile	Arg	Ile	Gly	Lys 510	Ala	Ser
Thr	Gly	Ser	Arg	Arg	Leu	Leu	Ser 520	Glu	Glu	Pro	Ser	Asp 525	Asp	Cys	Ile
Ser	Arg	Thr	Gln	Leu	Leu	Arg 535	Thr	Glu	Thr	Ala	Glu	Ile 540	His	Gly	Asp
Asn 545	Tyr	Gly	Gly	Pro	Gly 550	Asp	Lys	Ile	Thr	Ile 555	Cys	Asn	Gly	Ser	Thr 560
Ile	Val	Asp	Gln	Arg	Leu	Gly	Ser	Glu	Leu	Gly 570	Cys	Tyr	Thr	Ile 575	Asn
Arg	Val	Arg	Ser	Phe	Lys	Leu	Cys	Glu 585	Asn	Ser	Ala	Thr	Gly 590	Lys	Asn
Cys	Glu	Ile	Asp	Ser	Val	Pro	Val 600	Lys	Cys	Arg	Gln	Gly 605	Tyr	Cys	Leu
Arg	Ile	Thr	Gln	Glu	Gly	Arg 615	Gly	His	Val	Lys	Leu	Ser 620	Arg	Gly	Ser
Glu 625	Val	Val	Leu	Asp	Ala 630	Cys	Asp	Thr	Ser	Cys 635	Glu	Ile	Met	Ile	Pro 640
Lys	Gly	Thr	Gly	Asp 645	Ile	Leu	Val	Asp	Cys 650	Ser	Gly	Gly	Gln	Gln 655	His
Phe	Leu	Lys	Asp	Asn	Leu	Ile	Asp 665	Leu	Gly	Cys	Pro	Lys	Ile 670	Pro	Leu
Leu	Gly	Lys	Met	Ala	Ile	Tyr	Ile 680	Cys	Arg	Met	Ser	Asn 685	His	Pro	Lys
Thr	Thr	Met	Ala	Phe	Leu	Phe	Trp 695	Phe	Ser	Phe	Gly	Tyr 700	Val	Ile	Thr
Cys 705	Ile	Leu	Cys	Lys	Ala 710	Ile	Phe	Tyr	Leu	Leu 715	Ile	Ile	Val	Gly	Thr 720
Leu	Gly	Lys	Arg	Leu	Lys	Gln	Tyr	Arg	Glu	Leu 730	Lys	Pro	Gln	Thr 735	Cys
Thr	Ile	Cys	Glu	Thr	Thr	Pro	Val	Asn 745	Ala	Ile	Asp	Ala	Glu	Met	His 750
Asp	Leu	Asn	Cys	Ser	Tyr	Asn	Ile 760	Cys	Pro	Tyr	Cys	Ala	Ser	Arg	Leu 765

Thr 770	Ser	Asp	Gly	Leu	Ala	Arg 775	His	Val	Ile	Gln	Cys 780	Pro	Lys	Arg	Lys
Glu 785	Lys	Val	Glu	Glu	Thr 790	Glu	Leu	Tyr	Leu	Asn 795	Leu	Glu	Arg	Ile	Pro 800
Trp	Val	Val	Arg 805	Lys	Leu	Leu	Gln	Val	Ser 810	Glu	Ser	Thr	Gly	Val 815	Ala
Leu	Lys	Arg	Ser 820	Ser	Trp	Leu	Ile	Val 825	Leu	Leu	Val	Leu	Phe	Thr	Val
Ser	Leu	Ser	Pro	Val	Gln	Ser	Ala 840	Pro	Ile	Gly	Gln	Gly 845	Lys	Thr	Ile
Glu 850	Ala	Tyr	Arg	Ala	Arg	Glu 855	Gly	Tyr	Thr	Ser	Ile 860	Cys	Leu	Phe	Val
Leu 865	Gly	Ser	Ile	Leu	Phe 870	Ile	Val	Ser	Cys	Leu 875	Met	Lys	Gly	Leu	Val 880
Asp	Ser	Val	Gly 885	Asn	Ser	Phe	Phe	Pro	Gly 890	Leu	Ser	Ile	Cys	Lys 895	Thr
Cys	Ser	Ile	Ser 900	Ser	Ile	Asn	Gly	Phe 905	Glu	Ile	Glu	Ser	His 910	Lys	Cys
Tyr	Cys	Ser 915	Leu	Phe	Cys	Cys	Pro 920	Tyr	Cys	Arg	His 925	Cys	Ser	Thr	Asp
Lys 930	Glu	Ile	His	Lys	Leu	His 935	Leu	Ser	Ile	Cys	Lys 940	Lys	Arg	Lys	Lys
Gly 945	Ser	Asn	Val	Met	Leu 950	Ala	Val	Cys	Lys	Leu 955	Met	Cys	Phe	Arg	Ala 960
Thr	Met	Glu	Val 965	Ser	Asn	Arg	Ala	Leu	Phe 970	Ile	Arg	Ser	Ile	Ile	Asn 975
Thr	Thr	Phe 980	Val	Leu	Cys	Ile	Leu	Ile 985	Leu	Ala	Val	Cys	Val 990	Val	Ser
Thr	Ser 995	Ala	Val	Glu	Met	Glu	Asn 1000	Leu	Pro	Ala	Gly	Thr 1005	Trp	Glu	Arg
Glu 1010	Glu	Asp	Leu	Thr	Asn	Phe 1015	Cys	His	Gln	Glu	Cys 1020	Gln	Val	Thr	Glu
Thr 1025	Glu	Cys	Leu	Cys	Pro 1030	Tyr	Glu	Ala	Leu	Val 1035	Leu	Arg	Lys	Pro	Leu 1040
Phe	Leu	Asp	Ser 1045	Thr	Ala	Lys	Gly	Met	Lys 1050	Asn	Leu	Leu	Asn	Ser	Thr 1055
Ser	Leu	Glu	Thr 1060	Ser	Leu	Ser	Ile	Glu 1065	Ala	Pro	Trp	Gly	Ala 1070	Ile	Asn
Val	Gln 1075	Ser	Thr	Tyr	Lys	Pro	Thr 1080	Val	Ser	Thr	Ala	Asn 1085	Ile	Ala	Leu
Ser 1090	Trp	Ser	Ser	Val	Glu	His 1095	Arg	Gly	Asn	Lys	Ile 1100	Leu	Val	Ser	Gly
Arg 1105	Ser	Glu	Ser	Ile	Met 1110	Lys	Leu	Glu	Glu	Arg 1115	Thr	Gly	Ile	Ser	Trp 1120
Asp	Leu	Gly	Val 1125	Glu	Asp	Ala	Ser	Glu	Ser 1130	Lys	Leu	Leu	Thr	Val	Ser 1135
Val	Met	Asp 1140	Leu	Ser	Gln	Met	Tyr	Ser 1145	Pro	Val	Phe	Glu	Tyr 1150	Leu	Ser
Gly	Asp 1155	Arg	Gln	Val	Gly	Glu	Trp 1160	Pro	Lys	Ala	Thr	Cys 1165	Thr	Gly	Asp
Cys 1170	Pro	Glu	Arg	Cys	Gly 1175	Cys	Thr	Ser	Ser	Thr	Cys 1180	Leu	His	Lys	Glu
Trp	Pro	His	Ser	Arg	Asn	Trp	Arg	Cys	Asn	Pro	Thr	Trp	Cys	Trp	Gly

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1185	1190	1195	1200
Val Gly Thr Gly Cys Thr Cys Cys Gly Leu Asp Val Lys Asp Leu Phe	1205	1210	1215
Thr Asp Tyr Met Phe Val Lys Trp Lys Val Glu Tyr Ile Lys Thr Glu	1220	1225	1230
Ala Ile Val Cys Val Glu Leu Thr Ser Gln Glu Arg Gln Cys Ser Leu	1235	1240	1245
Ile Glu Ala Gly Thr Arg Phe Asn Leu Gly Pro Val Thr Ile Thr Leu	1250	1255	1260
Ser Glu Pro Arg Asn Ile Gln Gln Lys Leu Pro Pro Glu Ile Ile Thr	1265	1270	1275
Leu His Pro Arg Ile Glu Glu Gly Phe Phe Asp Leu Met His Val Gln	1285	1290	1295
Lys Val Leu Ser Ala Ser Thr Val Cys Lys Leu Gln Ser Cys Thr His	1300	1305	1310
Gly Val Pro Gly Asp Leu Gln Val Tyr His Ile Gly Asn Leu Leu Lys	1315	1320	1325
Gly Asp Lys Val Asn Gly His Leu Ile His Lys Ile Glu Pro His Phe	1330	1335	1340
Asn Thr Ser Trp Met Ser Trp Asp Gly Cys Asp Leu Asp Tyr Tyr Cys	1345	1350	1355
Asn Met Gly Asp Trp Pro Ser Cys Thr Tyr Thr Gly Val Thr Gln His	1365	1370	1375
Asn His Ala Ser Phe Val Asn Leu Leu Asn Ile Glu Thr Asp Tyr Thr	1380	1385	1390
Lys Asn Phe His Phe His Ser Lys Arg Val Thr Ala His Gly Asp Thr	1395	1400	1405
Pro Gln Leu Asp Leu Lys Ala Arg Pro Thr Tyr Gly Ala Gly Glu Ile	1410	1415	1420
Thr Val Leu Val Glu Val Ala Asp Met Glu Leu His Thr Lys Lys Ile	1425	1430	1435
Glu Ile Ser Gly Leu Lys Phe Ala Ser Leu Ala Cys Thr Gly Cys Tyr	1445	1450	1455
Ala Cys Ser Ser Gly Ile Ser Cys Lys Val Arg Ile His Val Asp Glu	1460	1465	1470
Pro Asp Glu Leu Thr Val His Val Lys Ser Asp Asp Pro Asp Val Val	1475	1480	1485
Ala Ala Ser Ser Ser Leu Met Ala Arg Lys Leu Glu Phe Gly Thr Asp	1490	1495	1500
Ser Thr Phe Lys Ala Phe Ser Ala Met Pro Lys Thr Ser Leu Cys Phe	1505	1510	1515
Tyr Ile Val Glu Arg Glu His Cys Lys Ser Cys Ser Glu Glu Asp Thr	1525	1530	1535
Lys Lys Cys Val Asn Thr Lys Leu Glu Gln Pro Gln Ser Ile Leu Ile	1540	1545	1550
Glu His Lys Gly Thr Ile Ile Gly Lys Gln Asn Ser Thr Cys Thr Ala	1555	1560	1565
Lys Ala Ser Cys Trp Leu Glu Ser Val Lys Ser Phe Phe Tyr Gly Leu	1570	1575	1580
Lys Asn Met Leu Ser Gly Ile Phe Gly Asn Val Phe Met Gly Ile Phe	1585	1590	1595
Leu Phe Leu Ala Pro Phe Ile Leu Leu Ile Leu Phe Phe Met Phe Gly	1605	1610	1615

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Trp Arg Ile Leu Phe Cys Phe Lys Cys Cys Arg Arg Thr Arg Gly Leu
 1620 1625 1630

Phe Lys Tyr Arg His Leu Lys Asp Asp Glu Glu Thr Gly Tyr Arg Arg
 1635 1640 1645

Ile Ile Glu Lys Leu Asn Asn Lys Lys Gly Lys Asn Lys Leu Leu Asp
 1650 1655 1660

Gly Glu Arg Leu Ala Asp Arg Arg Ile Ala Glu Leu Phe Ser Thr Lys
 1665 1670 1675 1680

Thr His Ile Gly

<210> SEQ ID NO 3
 <211> LENGTH: 482
 <212> TYPE: PRT
 <213> ORGANISM: Nairovirus
 <220> FEATURE:
 <223> OTHER INFORMATION: Nairovirus Crimean-Congo hemorrhagic fever
 virus (CCHFV) S (Small) protein, nucleocapsid protein

<400> SEQUENCE: 3

Met Glu Asn Lys Ile Glu Val Asn Asn Lys Asp Glu Met Asn Arg Trp
 1 5 10 15

Phe Glu Glu Phe Lys Lys Gly Asn Gly Leu Val Asp Thr Phe Thr Asn
 20 25 30

Ser Tyr Ser Phe Cys Glu Ser Val Pro Asn Leu Asp Arg Phe Val Phe
 35 40 45

Gln Met Ala Ser Ala Thr Asp Asp Ala Gln Lys Asp Ser Ile Tyr Ala
 50 55 60

Ser Ala Leu Val Glu Ala Thr Lys Phe Cys Ala Pro Ile Tyr Glu Cys
 65 70 75 80

Ala Trp Val Ser Ser Thr Gly Ile Val Lys Lys Gly Leu Glu Trp Phe
 85 90 95

Glu Lys Asn Ala Gly Thr Ile Lys Ser Trp Asp Glu Ser Tyr Thr Glu
 100 105 110

Leu Lys Val Asp Val Pro Lys Ile Glu Gln Leu Thr Gly Tyr Gln Gln
 115 120 125

Ala Ala Leu Lys Trp Arg Lys Asp Ile Gly Phe Arg Val Asn Ala Asn
 130 135 140

Thr Ala Ala Leu Ser Asn Lys Val Leu Ala Glu Tyr Lys Val Pro Gly
 145 150 155 160

Glu Ile Val Met Ser Val Lys Glu Met Leu Ser Asp Met Ile Arg Arg
 165 170 175

Arg Asn Leu Ile Leu Asn Arg Gly Gly Asp Glu Asn Pro Arg Gly Pro
 180 185 190

Val Ser His Glu His Val Asp Trp Cys Arg Glu Phe Val Lys Gly Lys
 195 200 205

Tyr Ile Met Ala Phe Asn Pro Pro Trp Gly Asp Ile Asn Lys Ser Gly
 210 215 220

Arg Ser Gly Ile Ala Leu Val Ala Thr Gly Leu Ala Lys Leu Ala Glu
 225 230 235 240

Thr Glu Gly Lys Gly Ile Phe Asp Glu Ala Lys Lys Thr Val Glu Ala
 245 250 255

Leu Asn Gly Tyr Leu Asp Lys His Lys Asp Glu Val Asp Arg Ala Ser
 260 265 270

Ala Asp Ser Met Ile Thr Asn Leu Leu Lys His Ile Ala Lys Ala Gln
 275 280 285

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Glu Leu Tyr Lys Asn Ser Ser Ala Leu Arg Ala Gln Ser Ala Gln Ile
 290 295 300
 Asp Thr Ala Phe Ser Ser Tyr Tyr Trp Leu Tyr Lys Ala Gly Val Thr
 305 310 315 320
 Pro Glu Thr Phe Pro Thr Val Ser Gln Phe Leu Phe Glu Leu Gly Lys
 325 330 335
 Gln Pro Arg Gly Thr Lys Lys Met Lys Lys Ala Leu Leu Ser Thr Pro
 340 345 350
 Met Lys Trp Gly Lys Lys Leu Tyr Glu Leu Phe Ala Asp Asp Ser Phe
 355 360 365
 Gln Gln Asn Arg Ile Tyr Met His Pro Ala Val Leu Thr Ala Gly Arg
 370 375 380
 Ile Ser Glu Met Gly Val Cys Phe Gly Thr Ile Pro Val Ala Asn Pro
 385 390 395 400
 Asp Asp Ala Ala Gln Gly Ser Gly His Thr Lys Ser Ile Leu Asn Leu
 405 410 415
 Arg Thr Asn Thr Glu Thr Asn Asn Pro Cys Ala Lys Thr Ile Val Lys
 420 425 430
 Leu Phe Glu Val Gln Lys Thr Gly Phe Asn Ile Gln Asp Met Asp Ile
 435 440 445
 Val Ala Ser Glu His Leu Leu His Gln Ser Leu Val Gly Lys Gln Ser
 450 455 460
 Pro Phe Gln Asn Ala Tyr Asn Val Lys Gly Asn Ala Thr Ser Ala Asn
 465 470 475 480
 Ile Ile

<210> SEQ ID NO 4
 <211> LENGTH: 7
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Crimean-Congo hemorrhagic
 fever virus (CCHFV) Afg09-2990 strain L (Large)
 protein conserved region

<400> SEQUENCE: 4

Trp Thr Gln Val Ile Asp Gly
 1 5

<210> SEQ ID NO 5
 <211> LENGTH: 7
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Crimean-Congo hemorrhagic
 fever virus (CCHFV) Afg09-2990, Matin, Oman,
 Baghdad, UG3010, Turkey and SPU128-81 strains L
 (Large) protein conserved region

<400> SEQUENCE: 5

Glu Glu Pro Glu Ala Lys Leu
 1 5

<210> SEQ ID NO 6
 <211> LENGTH: 7
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Crimean-Congo hemorrhagic
 fever virus (CCHFV) Matin strain L (Large) protein
 conserved region

-continued

<400> SEQUENCE: 6

Trp Thr Gln Val Ile Asp Ser
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<210> SEQ ID NO 7

<211> LENGTH: 7

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic Nairovirus Crimean-Congo hemorrhagic
 fever virus (CCHFV) Oman, UG3010, IBAr10200 and
 SPU128-81 strains L (Large) protein conserved
 region

<400> SEQUENCE: 7

Trp Thr Gln Val Ile Ala Gly
 1 5

<210> SEQ ID NO 8

<211> LENGTH: 7

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic Nairovirus Crimean-Congo hemorrhagic
 fever virus (CCHFV) Baghdad strain L (Large)
 protein conserved region

<400> SEQUENCE: 8

Trp Thr Gln Val Met Ala Gly
 1 5

<210> SEQ ID NO 9

<211> LENGTH: 7

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic Nairovirus Crimean-Congo hemorrhagic
 fever virus (CCHFV) Turkey strain L (Large)
 protein conserved region

<400> SEQUENCE: 9

Trp Thr Gln Val Ile Ala Ser
 1 5

<210> SEQ ID NO 10

<211> LENGTH: 7

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic Nairovirus Crimean-Congo hemorrhagic
 fever virus (CCHFV) IBAr10200 strain L (Large)
 protein conserved region

<400> SEQUENCE: 10

Glu Glu Pro Glu Ala Arg Leu
 1 5

<210> SEQ ID NO 11

<211> LENGTH: 7

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic Nairovirus Crimean-Congo hemorrhagic
 fever virus (CCHFV) AP92Greece strain L (Large)
 protein conserved region

<400> SEQUENCE: 11

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Trp Thr Gln Val Leu Ala Gly
1 5

<210> SEQ ID NO 12
 <211> LENGTH: 7
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Crimean-Congo hemorrhagic
 fever virus (CCHFV) AP92Greece strain L (Large)
 protein conserved region

<400> SEQUENCE: 12

Asp Glu Pro Glu Ala Lys Leu
1 5

<210> SEQ ID NO 13
 <211> LENGTH: 7
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Dugbe virus (DUGV) L
 (Large) protein conserved region

<400> SEQUENCE: 13

Trp Glu Arg Val Val Asp Glu
1 5

<210> SEQ ID NO 14
 <211> LENGTH: 8
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Dugbe virus (DUGV) L
 (Large) protein conserved region

<400> SEQUENCE: 14

Thr Glu Pro Glu Ala Val Gly Thr
1 5

<210> SEQ ID NO 15
 <211> LENGTH: 8
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Crimean-Congo hemorrhagic
 fever virus (CCHFV) L (Large) protein conserved
 region

<400> SEQUENCE: 15

Glu Glu Pro Glu Ala Arg Leu Val
1 5

<210> SEQ ID NO 16
 <211> LENGTH: 7
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Nairobi sheep disease
 virus (NSDV) L (Large) protein conserved region

<400> SEQUENCE: 16

Trp Glu Glu Val Val Pro Gly
1 5

<210> SEQ ID NO 17
 <211> LENGTH: 8
 <212> TYPE: PRT

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<213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Nairobi sheep disease
 virus (NSDV) L (Large) protein conserved region

 <400> SEQUENCE: 17

 Glu Glu Pro Glu Ala Lys Gly Leu
 1 5

 <210> SEQ ID NO 18
 <211> LENGTH: 6
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Erve virus (ERVEV) L
 (Large) protein conserved region

 <400> SEQUENCE: 18

 Trp Glu Asn Ile Glu Gly
 1 5

 <210> SEQ ID NO 19
 <211> LENGTH: 8
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Erve virus (ERVEV) L
 (Large) protein conserved region

 <400> SEQUENCE: 19

 Ile Glu Pro Glu Ala Ile Gly Leu
 1 5

 <210> SEQ ID NO 20
 <211> LENGTH: 7
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Hazara virus (HAZV) L
 (Large) protein conserved region

 <400> SEQUENCE: 20

 Trp Asp Ser Val Ser Asp Ile
 1 5

 <210> SEQ ID NO 21
 <211> LENGTH: 8
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic Nairovirus Hazara virus (HAZV) L
 (Large) protein conserved region

 <400> SEQUENCE: 21

 Thr Glu Pro Glu Ala Ala Ala Thr
 1 5

 <210> SEQ ID NO 22
 <211> LENGTH: 6
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic ubiquitin substrate carboxy terminal
 of ubiquitin hexapeptide homolog
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (6)...(6)
 <223> OTHER INFORMATION: Gly modified by 7-amino-4-methylcoumarin (AMC)
 fluorescent tag

-continued

<400> SEQUENCE: 22

Glx Arg Leu Arg Gly Gly
 1 5

<210> SEQ ID NO 23

<211> LENGTH: 6

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic ubiquitin substrate carboxy terminal
 of ubuquitin hexapeptide homolog

<400> SEQUENCE: 23

Glx Arg Leu Arg Gly Gly
 1 5

The invention claimed is:

1. A pharmaceutical composition effective in eliciting a specific immune response, comprising a recombinantly altered Crimean-Congo hemorrhagic fever (CCHF) virus comprising an L protein that has been recombinantly altered to have decreased deubiquitinating activity or decreased deISGylating activity while maintaining protease activity, such that the CCHF virus replicates in human cells, wherein the recombinantly altered L protein is altered at a position corresponding to a ubiquitin or ISG15 substrate binding interface of OTU domain protease.

2. The immunogenic composition of claim 1, which has been recombinantly altered to have both decreased deubiquitinating activity and decreased deISGylating activity.

3. The immunogenic composition of claim 1, wherein the virus has been chemically or radiologically inactivated.

4. The immunogenic composition of claim 1, which has been modified wherein the L protein comprises a substitution at position 13, position 77, or both position 13 and 77 of the L protein.

5. The immunogenic composition of claim 4, wherein position 13 of the L protein is changed to arginine.

6. The immunogenic composition of claim 4, wherein position 77 of the L protein is changed to aspartic acid.

7. The immunogenic composition of claim 1, further comprising an adjuvant.

8. A recombinantly altered CCHF virus comprising an L protein that has been recombinantly altered to have both decreased deubiquitinating activity and decreased deISGylating activity while maintaining protease activity, such that the CCHF virus replicates in human cells, wherein the recombinantly altered L protein is altered at a position corresponding to a ubiquitin or ISG15 substrate binding interface of OTU domain protease.

9. The recombinantly altered virus of claim 8, which has been modified wherein the L protein comprises a substitution at position 13, position 77, or both position 13 and 77 of the L protein.

10. The recombinantly altered virus of claim 8, wherein position 13 of the L protein is changed to arginine.

11. The recombinantly altered virus of claim 8, wherein position 77 of the L protein is changed to aspartic acid.

12. The recombinantly altered virus of claim 8, wherein the virus has no ability or a reduced ability to inhibit expression of interferon β .

13. A host human cell line transfected with a recombinantly altered virus according to claim 8.

14. A method of eliciting an immune response against a recombinantly altered CCHF virus, comprising administering to a subject in need thereof an immunogenic composition according to claim 1.

15. A method of developing an immunogenic but substantially non-pathogenic CCHF virus, comprising:

- a) transfecting a host cell with the genome of a recombinantly altered CCHF virus;
- b) transfecting the host cell with a codon optimized L protein expression vector and an N protein expression vector;
- c) obtaining replicated virus particles comprising said genetic alterations from the host cell; and
- d) testing the replicated virus particles for decreased deubiquitinating activity and/or decreased deISGylating activity; and
- e) selecting one or more virus particles with decreased deubiquitinating activity and/or decreased deISGylating activity.

16. The method of claim 15, wherein step (a) comprises transfecting the host cell with the L, M, and S gene sectors in separate vectors.

17. A method for preparing a commercial product, comprising packaging a pharmaceutical composition according to claim 1 with information on how to use the product for eliciting an immune response against a CCHF virus.

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